



## REVIEW ARTICLE

# Critical evaluation of key evidence on the human health hazards of exposure to bisphenol A

J. G. Hengstler,<sup>1</sup> H. Foth,<sup>2</sup> T. Gebel,<sup>3</sup> P.-J. Kramer,<sup>4</sup> W. Lilienblum,<sup>5</sup> H. Schweinfurth,<sup>6</sup> W. Völkel,<sup>7</sup> K.-M. Wollin,<sup>8</sup> and U. Gundert-Remy<sup>9</sup>

<sup>1</sup>Leibniz Research Centre for Working Environment and Human Factors (IfADo), University of Dortmund, Dortmund, Germany, <sup>2</sup>Institute of Environmental Toxicology, University of Halle, Halle/Saale, Germany, <sup>3</sup>Federal Institute for Occupational Safety and Health, Dortmund, Germany, <sup>4</sup>Research and Development, Merck Serono, Darmstadt, Germany, <sup>5</sup>Dr. Lilienblum Consulting Toxicology LiCoTox, Hemmingen/Han, Germany, <sup>6</sup>Global Early Development, Bayer Schering Pharma AG, Berlin, Germany, <sup>7</sup>Bavarian Health and Food Safety Authority, Munich, Germany, <sup>8</sup>Lower Saxony Governmental Institute of Public Health, Hannover, Germany, and <sup>9</sup>Federal Institute for Risk Assessment (BfR), Berlin, Germany

---

### Abstract

Despite the fact that more than 5000 safety-related studies have been published on bisphenol A (BPA), there seems to be no resolution of the apparently deadlocked controversy as to whether exposure of the general population to BPA causes adverse effects due to its estrogenicity. Therefore, the Advisory Committee of the German Society of Toxicology reviewed the background and cutting-edge topics of this BPA controversy. The current tolerable daily intake value (TDI) of 0.05 mg/kg body weight [bw]/day, derived by the European Food Safety Authority (EFSA), is mainly based on body weight changes in two- and three-generation studies in mice and rats. Recently, these studies and the derivation of the TDI have been criticized. After having carefully considered all arguments, the Committee had to conclude that the criticism was scientifically not justified; moreover, recently published additional data further support the reliability of the two- and three-generation studies demonstrating a lack of estrogen-dependent effects at and below doses on which the current TDI is based. A frequently discussed topic is whether doses below 5 mg/kg bw/day may cause adverse health effects in laboratory animals. Meanwhile, it has become clear that positive results from some explorative studies have not been confirmed in subsequent studies with higher numbers of animals or a priori defined hypotheses. Particularly relevant are some recent studies with negative outcomes that addressed effects of BPA on the brain, behavior, and the prostate in rodents for extrapolation to the human situation. The Committee came to the conclusion that rodent data can well be used as a basis for human risk evaluation. Currently published conjectures that rats are insensitive to estrogens compared to humans can be refuted. Data from toxicokinetics studies show that the half-life of BPA in adult human subjects is less than 2 hours and BPA is completely recovered in urine as BPA-conjugates. Tissue deconjugation of BPA-glucuronide and -sulfate may occur. Because of the extremely low quantities, it is only of minor relevance for BPA toxicity. Biomonitoring studies have been used to estimate human BPA exposure and show that the daily intake of BPA is far below the TDI for the general population. Further topics addressed in this article include reasons why some studies on BPA are not reproducible; the relevance of oral versus non-oral exposure routes; the degree to which newborns are at higher systemic BPA exposure; increased BPA exposure by infusions in intensive care units; mechanisms of action other than estrogen receptor activation; and the current regulatory status in Europe, as well as in the USA, Canada, Japan, New Zealand, and Australia. Overall, the Committee concluded that the current TDI for BPA is adequately justified and that the available evidence indicates that BPA exposure represents no noteworthy risk to the health of the human population, including newborns and babies.

**Keywords:** Biomonitoring, endocrine disruption, estrogenic effects, intensive care units, interspecies extrapolation, low-dose effects, regulation, three- and two-generation studies, tissue deconjugation, tolerable daily intake, toxicokinetic modeling

---

*Address for Correspondence:* J. G. Hengstler, Leibniz Research Centre for Working Environment and Human Factors (IfADo), University of Dortmund, Ardeystrasse 67, 44139 Dortmund, Germany. E-mail: hengstler@ifado.de; U. Gundert-Remy, Federal Institute for Risk Assessment (BfR), 14195 Berlin, Germany. E-mail: gundert@googlemail.com

(Received 05 November 2010; revised 19 January 2011; accepted 25 January 2011)

## Contents

Abstract .....	263
Introduction.....	264
Derivation of tolerable daily intake values.....	265
Are the three- and two-generation studies of Tyl et al. valid?.....	265
Do oral low BPA doses below 5 mg/kg bw/day cause adverse health effects in laboratory animals? .....	267
How can differences between industry sponsored and publicly sponsored studies be explained? .....	270
Toxicokinetics .....	271
Humans.....	271
Non-human primates .....	275
Rats .....	275
Mice .....	275
Importance of the exposure route.....	275
Can rodents be used to extrapolate to the human situation with respect to estrogenic activity?.....	276
Are there susceptible subpopulations?.....	277
Toxicokinetics in children.....	277
Toxicokinetics in other groups .....	278
Tissue deconjugation of BPA-glucuronide and BPA-sulfate .....	278
Toxicodynamics.....	279
Specific exposure conditions .....	279
Neonatal intensive care unit.....	279
Polycarbonate (PC) bottles.....	279
How can biomonitoring support risk evaluation? .....	280
Results and interpretation of biomonitoring studies with BPA .....	280
Biomonitoring and exposure assessment .....	281
What are the mechanisms of action of BPA? Does the multitude of mechanisms besides estrogen receptor activation make the substance more hazardous?.....	282
Epidemiological studies in the general population .....	283
Recent governmental responses .....	283
European Union .....	283
Denmark .....	284
France.....	284
Switzerland .....	284
Australia and New Zealand.....	284
Canada .....	285
USA .....	285
Japan.....	285
Acknowledgments .....	285
Declaration of interest.....	286
References.....	286

## Introduction

For more than 10 years there has been a scientific and journalistic controversy whether bisphenol A (BPA) causes adverse effects in humans related to its estrogenic activity (Borrell, 2010; Lorentzen and Hattan, 2010; Aschberger et al., 2010; Taylor et al., 2010; Yang et al., 2009). BPA, a building block of polycarbonate plastic and epoxy resins, was first synthesized in 1891. However, commercial production did not begin before the early 1950s when the first epoxy resins were developed (Vogel, 2009). Epoxy resins are extensively used as protective coatings on metal equipment, food cans, piping, and dental sealants. In 1957, it was discovered that

polymerization of BPA with phosgene leads to polycarbonate. There is an unusual wealth of safety-related studies carried out on BPA. These cover nearly every possible endpoint. Its estrogenic properties were described as early as 1936 (Dodds and Lawson, 1936). To date, more than 5000 studies on BPA have been published. It is obvious that this should be enough information to resolve the controversy, but nevertheless this has not yet been achieved and those not directly involved in BPA research are usually puzzled by the never-ending and sometimes emotional debate. In order to contribute to a balanced and well-founded resolution of the seemingly deadlocked situation, the Advisory Committee of the German

Society of Toxicology<sup>1</sup> reviewed the background and the cutting-edge questions of the BPA controversy (Table 1) and offers an independent judgement.

### Derivation of tolerable daily intake values

BPA has been tested in subchronic oral toxicity studies using rats, mice, and dogs (US EPA, 1984 a, 1984b, 1984c; NTP, 1982). Rats and mice were administered BPA in the diet for 90 days (250–4000 ppm in rats; 5000–25,000 ppm in mice) (NTP, 1982). Doses higher than 1000 ppm (equivalent to approximately 67 mg/kg/day) lead to decreased body weight in both sexes of rats. In dogs (90 days; 1000–9000 ppm BPA in the diet), the only toxic effect observed was an increase in mean liver weight in the high-dose group (US EPA, 1984a). Bisphenol A was evaluated for developmental toxicity in CD rats (0, 160, 320, or 640 mg/kg bw/day) and CD-1 mice (0, 500, 750, 1000, or 1250 mg/kg bw/day) dosed daily by gastric intubation from gestational days 6 through 15. In Charles River rats, the only effect observed in two-generation studies (100–9000 ppm BPA in the diet) was decreased body weight in the F0 generation at 9000 ppm and in the F1 generation at doses equal or higher than 1000 ppm (US EPA, 1984b, 1984c). Also, male mice receiving doses higher than 15,000 ppm and the exposed females exhibited a decreased body weight gain compared to the controls (in mice 15,000 ppm is equivalent to approximately 1950 mg/kg/day based on a food factor of 0.13). In mice, doses of 1250 mg/kg/day led to maternal toxicity, including fetotoxic effects. However, no significant increase in the incidence of malformations was observed (NTP, 1985). In rats, doses equal to and higher than 1280 mg/kg/day were not toxic and did not cause malformations of the fetus (NTP, 1986).

Taken together, the toxic effects observed in laboratory animals after repeated BPA exposure occur at doses that are several magnitudes higher than the exposure of the general human population. Since it is not clear to which extent these rather early studies mentioned above accounted for internal quality assurance and were conducted according to accepted modern testing guidelines (or elements of them), their validity, reliability, and thus value for hazard and risk assessment are considered limited. Modern testing guidelines represent internationally agreed-upon guidance with the goal to provide a reproducible set of data that can optimally satisfy all internationally agreed safety assessment criteria used in

the regulatory processes. Hence, the focus of this article is on findings of more recent studies conducted for regulatory purposes.

In a three-generation rat study (Tyl et al., 2002) and a two-generation study in mice (Tyl et al., 2008b, 2008c), BPA was found to decrease body weight, and the weights of the livers and kidneys. An overall no observed adverse effect level (NOAEL) of 5 mg BPA/kg bw/day was derived, based on liver weight decreases, the most sensitive endpoint. At doses leading to liver weight changes, BPA did not cause any effects on hormone-sensitive endpoints, which are the focus of concern of the current debate on BPA toxicity. Using an uncertainty factor of 100 (10 for interspecies differences, 10 for interindividual differences), a tolerable daily intake (TDI) of 0.05 mg/kg bw/day was set by the European Food Safety Authority (EFSA, 2006) and confirmed in 2008 and 2010 (EFSA, 2008, 2010). This TDI has been accepted by most regulatory agencies worldwide. Similarly, the United States Environmental Protection Agency (US EPA) derived a reference dose of 0.05 mg/kg bw/day (reviewed by Willhite et al., 2008). Perhaps the most intensively currently discussed topic is whether the NOAELs used for deriving the TDI are scientifically valid and appropriate for risk assessment.

### Are the three- and two-generation studies of Tyl et al. valid?

The studies of Tyl et al. (2002, 2008b, 2008c) did not reveal any effects on fertility or development. Doses of 0.001, 0.02, 0.3, 5, 50, and 500 mg/kg bw/day of BPA were tested in CD Sprague-Dawley rats in a three-generation study (Tyl et al., 2002), and 0.003, 0.03, 0.3, 5, 50, as well as 600 mg BPA/kg bw/day in CD-1 mice in a two-generation study (Tyl et al., 2008b). The dose ranges in the latter studies also cover the “low-dose range.” Another two-generation reproductive toxicity study performed under good laboratory practice (GLP) did also not

Table 1. Cutting edge topics of the current controversy on BPA.

Are the studies used for regulatory purposes flawed?
Do oral low doses below 5 mg/kg bw/day cause adverse health effects in laboratory animals?
How can differences between industry sponsored and publicly sponsored studies be explained?
Swallow or inject? What is the relevance of the oral route versus implantation of pumps or intravenous injection?
Can rodents be used to extrapolate to the human situation?
To which degree are embryos, babies, or children more susceptible?
How critical is exposure by intravenous infusion in intensive care units?
How critical is tissue deconjugation of BPA-glucuronide and BPA-sulfate?
How can biomonitoring support risk evaluation?
What are the mechanisms of action of BPA? Does the multitude of mechanisms other than estrogen receptor activation make the substance more dangerous?
Why are recent governmental responses inconsistent?

<sup>1</sup> The Advisory Committee of the German Society of Toxicology is elected by the members of the German Society of Toxicology and consists of representatives from academia, industry, and administration in order to guarantee a broad range of toxicological competence. The Advisory Committee may consult further experts with expertise in specific fields of Toxicology. In case of the present work, the committee included Dr. Wolfgang Völkel as an additional expert and sent the manuscript to Wolfgang Dekant, and Regine Kahl for discussion. The Advisory Committee presents and justifies its activities to the members of the German Society of Toxicology, for example at the yearly plenary meeting. The German Society of Toxicology is the largest scientific toxicological organization in Europe, with more than 1000 members..

Table 2. Criticisms of the studies of Tyl et al. (2008a, 2008b, 2008c) and responses.

Criticism	Response
1. Myers et al. questioned the use of CD-1 Swiss mice because of their "aberrant insensitivity" to estrogens.	Recently, Ryan et al. (2010a) used a wide range of doses from 0.05 to 50 µg/kg bw/day for the positive control estrogen, EE2. They observed a comparable sensitivity for CD-1 mice, Long-Evans, and Sprague-Dawley rats for several adverse effects. Therefore, the use of CD-1 mice by Tyl et al. (2008a, 2008b, 2008c) is justified. Tyl et al. purchased their CD-1 mice from Charles River to avoid supplier effects. It should be considered that also vom Saal and some of his colleagues used CD-1 mice and reported low dose effects (Timms et al., 2005). Low-dose effects of BPA were also reported in Long-Evans (Akingbemi et al., 2004) and in SD rats.
2. Myers et al. considered the prostate weights reported in the study of Tyl et al. (2008a, 2008b, 2008c) "abnormally high" and suggested "that the dissection procedures for the prostate in the Tyl laboratory included other nonprostatic tissues in the weight measurements, rendering them unusable..." This criticism was also expressed by Gies et al. (2009).	Tyl (2009a, 2009b) reported that paraffin blocks and slides of the analyzed prostates are still available and indicate no evidence of extraneous tissue/fat or excessive inflammation (all recently audited by the US FDA). In a response letter (Tyl, 2009a, 2009b), the author documented that considering the age of the mice, prostate weights in their study were within the range of published data. Differences from vom Saal's study may be explained by his post wean caging regimen. Vom Saal's CF-1 mice were housed by sex. Pups were weaned at 23 days of age and male littermates were housed three per cage (Nagel et al., 1997). Prostate weight was determined when the mice were 6 months old. For this purpose, one male per litter was randomly selected and individually housed for 1 month before determination of prostate weights (Nagel et al., 1997). This procedure is problematic because the dominant cage male develops large androgen-dependent accessory organs, whereas subservient cage males have smaller prostates. It is questionable whether 1 month of single housing may compensate for months of group housing and its influence on sexual development of male mice. In the study of vom Saal (Nagel et al., 1997), no controls were presented with respect to the influence of group housing and compensation by single housing and the authors did not report whether they determined prostate weight only from the dominant males or if all animals were included. Since the authors wrote that "one male per litter was randomly selected..." (Nagel et al., 1997), they likely included both, dominant and subservient males. This may explain the high variability of the prostate weights in studies from vom Saal's lab compared to much lower variability in Tyl's studies (Tables 1 and 2 in Tyl, 2009a, 2009b). It should also be taken into account that the number of animals exposed to BPA in vom Saal's study ( $n=7$ ) is very low also, considering that an additional confounder (dominant versus subservient mice) may be present. Much higher numbers of animals were used in Tyl's studies (28 per group). After careful evaluation of Tyl's studies, the criticism of Myers et al. (2009) and Gies et al. (2009) concerning prostate weights is unsubstantiated. However, several aspects may be critical in vom Saal's studies, such as the influence of dominant versus subservient males, no available histology, and use of a now obsolete in-house strain preventing other researchers to reproduce the initial positive studies. Another drawback is the statistical analysis. Concerning the criticism that crude mistakes were made by Tyl's laboratory in determining prostate weights, it should also be considered that this laboratory has carried out a large number of reprotox studies (often sponsored by the US Government) and has successfully joined interlaboratory validation studies, where the Tyl laboratory achieved some of the most precise prostate weight data and robust treatment-related effects (EPA review).
3. Myers et al. criticized the high incidence and severity of prostatitis in the animals from Tyl's study, which may compromise their results.	The paraffin slides of the prostates of the mice from Tyl's study are still available and have been reanalyzed by an independent pathologist. The results have been published (Tyl, 2009a, 2009b). The incidence of inflammation in CD-1 mice of Tyl's study matches the low incidences of prostatitis seen in many mouse strains without treatment. Therefore, it is extremely unlikely that the results of Tyl et al. (2008a, 2008b, 2008c) have been compromised by prostatitis. It should be kept in mind that vom Saal's laboratory did not include any histopathological examinations (Nagel et al., 1997). Therefore, a possible confounding influence of prostatitis could not be accounted for in their study.
4. Myers et al. criticized that the diet used in Tyl's study contained phytoestrogens, which they claim would interfere with BPA activity.	Recently, Tyl et al. have published the genistein, daidzein, and glycitein contents of the standard diet (Purina Certified Ground Rodent Chow No. 5002) used in their study. Vom Saal and colleagues have not reported the phytoestrogen content of their diet. The majority of "normal rodent diets have similar levels of phytoestrogens." Although it is not possible to compare diets between Tyl's and vom Saal's laboratories, it is extremely unlikely that the diet in Tyl's study compromised an estrogenic response, because in studies with estradiol in which mice were fed the same standard diet as those in the BPA studies, estradiol (0.5 ppm) clearly accelerated acquisition of puberty (Tyl, 2008a).



Table 2. Continued.

Criticism	Response
5. Myers et al. criticized that “Tyl et al. (2008a) did not examine any neurobehavioral end points,” which Myers et al. interpreted as a “glaring omission of Tyl.”	The multigeneration study performed by Tyl et al. (2008a) represents a standard test performed according to a specific guideline that is not designed to investigate neurobehavioral endpoints. Therefore, there is certainly no “glaring omission of Tyl,” because behavior simply is not an endpoint in this study type. However, there are other studies that have focussed on neurobehavioral endpoints: Ema et al. (2001) observed no effects in their two-generation, low-dose (0.2–200 µg BPA/kg bw/day) study. Similarly, Ryan et al. (2010a) obtained negative results in a detailed study on reproductive development, function, and behavior in female rats exposed perinatally to a wide range of BPA doses (see also Sharpe, 2010). Stump et al. (2010) published the most recent study conducted according to GLP. The results show that BPA, at levels of exposure 4000-fold higher than the maximum human exposure in the general population, does not cause any discernible adverse effects in female rats. By contrast, EE2, used as a positive control in this study, caused major adverse health effects at doses in the range of those applied in early contraceptives (Ryan et al., 2010a; highlight report: Sharpe, 2010).
6. Myers et al. criticized Tyl et al. (2008a, 2008b, 2008c) for using too many animals: “...all of the studies by Tyl et al. were significantly overpowered and this is in direct violation of federal guidelines for conducting animal research, a fact about which U.S. FDA regulators seem unaware.”	The consideration of animal protection aspects should certainly be an important point in all animal testing. But even if we assume that more animals than required were used, according to statistical power analysis, this would not lead to different or incorrect conclusions from a study. However, the numbers of animals used in Tyl’s studies are appropriate—28 mice/sex/group/generation. This is in line with the OECD and US EPA toxicity testing guidelines, which require at least 20 pregnant females per group.

observe effects in the low-dose range (0.2–200 µg/kg bw/day) (Ema et al., 2001). However, criticism has been raised, e.g., by Myers et al. (2009), who described the studies of Tyl et al. (2002, 2008; Tyl 2009a, 2009b) as “so flawed as to be useless.” If these studies are considered invalid, this would have serious consequences indeed. Therefore, we collected the points raised by the critics and carefully considered their relevance. Considering the arguments summarized in Table 2, we came to the conclusion that all criticisms were already refuted convincingly by the author (R. Tyl) herself. In this context, it should also be considered that a further study published recently has confirmed a complete absence of effects over a wide range of BPA exposures in female rats exposed perinatally (Ryan et al., 2010a). In this study, BPA exposure 40-, 400-, or even 4000-fold higher than the maximum estimated exposure to humans in the general population caused no adverse effects (Ryan et al., 2010a; Sharpe, 2010). The endpoints in this study were adult sex hormone-dependent behavior and female reproductive development. The results are consistent with previously published studies that have shown an absence of reproductive effects in male rats and mice (for example: Tyl et al., 2002; Ema et al., 2001; Tinwell et al., 2002).

The argument of Myers et al. (2009), claiming that estrogen-insensitive mouse and rat strains have been used by Tyl et al (2002, 2008b, 2008c), has to be judged as not valid. As pointed out by Ryan et al. (2010b), CD-1 mice respond to low doses of exogenously administered estrogens, as do the rat strains used by these authors (Ryan et al., 2010b). Nagel et al. (1997), in vom Saal’s laboratory, reported enlarged prostates in F1 adult CF-1 mice from in utero exposure to 2 and 20 µg/kg bw/day oral BPA. These results could not be confirmed by other investigators who tried to reproduce the study with

higher numbers of animals (Cagen et al., 1999; Ashby et al., 1999). Unfortunately, vom Saal’s laboratory has meanwhile closed their colony of CF-1 mice, which precludes reproduction of the study. Therefore, the question remains open as to whether this strain was particularly sensitive to BPA or whether the positive result was an artefact.

We addressed the criticism of Gies et al. (2009) that “Also no reason was found why the GLP-studies (Tyl et al., 2002, 2008; Tyl, 2003) used for regulatory purposes (by European authorities) did not find BPA effects and whether the reported prostate weights are inconsistent with literature data.” The most obvious explanation for the lack of BPA effects is that the compound simply does not cause any adverse health effects in the low-dose range. Also, the prostate weights reported by Tyl et al. (2002, 2008) are certainly not inconsistent with literature data. Considering age and caging schedules, the prostate weights of interest are in agreement with published literature (Tables 1 and 2 in Tyl, 2009a, 2009b). Data have been obtained in a commercially available mouse strain and can therefore easily be reproduced.

In conclusion, the criticism of Myers et al. (2009) and Gies et al. (2009; see also comments to this paper at: <http://www.umweltbundesamt.de/gesundheit-e/veranstaltungen/bisphenol-a/index.htm>) regarding the value of the three- and two-generation studies of Tyl et al. are unsubstantiated.

### **Do oral low BPA doses below 5 mg/kg bw/day cause adverse health effects in laboratory animals?**

Several guideline-compliant toxicity studies have been performed that resulted in a systemic NOAEL of 5 mg BPA/kg bw/day and a reproductive/developmental NOAEL

of 50 mg BPA/kg bw/day in rats and mice (Table 3). Consistently, no adverse health effects were observed at doses  $\leq$  5 mg BPA/kg/day.

In addition to these guideline-compliant studies, a high number of exploratory research studies have been performed that usually study early, molecular, or other endpoints, of which the relevance for adverse health effects often has not yet been validated. If these studies are reproducible and their interpretation is clear, they should be used to support risk assessment. However, conclusions from explorative research studies on BPA were inconsistent (reviewed in Goodman et al., 2006; 2009; Gray et al., 2004; Willhite et al., 2008; Chapin et al., 2008; EFSA, 2006). For example, vom Saal's laboratory performed a study in F1 adult mouse offspring (CF-1 mice originally obtained from Charles River and later maintained as an outbred colony in the authors' facility), which were orally dosed with BPA at 2 and 20  $\mu$ g/kg bw/day on gestational days 11–17 (Nagel et al., 1997). The authors reported similar increases in prostate weights in both exposure groups (2 and 20  $\mu$ g/kg bw/day) of 30–35% compared to controls. However, this result could not be reproduced in later studies with higher statistical power, for example by Cagen et al. (1999) and Ashby et al. (1999). The negative outcomes in the studies of Cagen et al. (1999) and Ashby et al. (1999) were later criticized by vom Saal and colleagues (Myers et al., 2009), because they were not considered useful for risk assessment by the NTP-CERHR (National Toxicology Program Center for the Evaluation of Risks to Human Reproduction). However, the studies of Cagen et al. (1999) and Ashby et al. (1999) had been designed to repeat and verify vom Saal's study with larger numbers of animals. They were not intended for regulatory purposes and, therefore, did not follow the protocol of a toxicity testing guideline. Possible reasons for the non-reproducibility of vom Saal's study are the small numbers of only seven mice per dose groups and the lack of control of the confounding effect of dominant versus subservient mice on prostate weights during group housing (Table 2, criticism 2).

In a situation with apparently contradictory data, it is prudent to have the complete set of data evaluated by a panel of experts (Chapin et al., 2008; EFSA, 2006, 2008, 2010 a, 2010b; Goodman et al., 2006, 2009; US FDA Memorandum, 2009a, 2009b; Willhite et al., 2008). Therefore, the NTP-CERHR formed such an expert panel, including several internationally known scientists working on BPA, toxicologists, epidemiologists, and statisticians (NTP, 2008). This expert panel has evaluated more than 700 studies trying to extract possible evidence of adverse health effects. Many of the studies included in the assessment failed to meet minimal quality criteria for experimental design and statistical analysis.<sup>2</sup> Among the most common deficiencies was a failure to control for "litter effects," although litter-based statistics have

been specified by the US EPA. Siblings from the same litter often have similar properties. If this is not considered in the study design, random variation between litters may be misinterpreted as a signal of a treatment-related effect. The expert panel also did not consider studies that did not include a concurrent control group of animals, injected BPA into the brain or spinal cord, or contained positive control groups that did not show adverse effects. Finally, the NTP expert panel applied the five possible levels of concern (negligible concern, minimal concern, some concern, concern, and serious concern) and concluded that there was "some concern" for BPA-associated effects on the brain, behavior, and prostate, whereas most other effects were rated as of "negligible" or "minimal" concern (NTP, 2008). Consequently, further studies were designed to clarify the situation where the NTP expert panel expressed "some concern." One example is the study by Ryan et al. (2010a) in which pregnant Long-Evans rats were treated orally by gavage with 0, 2, 20, and 200  $\mu$ g BPA/kg bw/day from day 7 of gestation to postnatal day 18. In the female offspring that were examined, BPA did not alter sexually dimorphic behavior, puberty, fertility, or anatomy (Ryan et al., 2010a). In the same study, ethinyl estradiol (EE2) at doses of 0.05–50  $\mu$ g/kg bw/day increased anogenital distance, reduced pup body weight, accelerated age at vaginal opening, reduced F1 fertility and F2 litter sizes, and induced malformations of the external genitalia (Ryan et al., 2010a). In a second study, BPA (2, 20, or 200  $\mu$ g/kg bw/day) was administered by oral gavage during pregnancy from gestational day 7 to postnatal day 18 and the male offspring were studied (Howdeshell et al., 2008). BPA did not affect androgen-dependent reproductive organ weights, including prostate weights or epididymal sperm abundance. By contrast, adult body weight was reduced by EE2 at 50  $\mu$ g/kg bw/day and androgen-dependent tissue weights were reduced in a dose-dependent manner (Howdeshell et al., 2008). Therefore, these new studies have addressed some of the endpoints about which the NTP expert panel had expressed "some concern." The negative outcome of these large and well-designed studies prompted the question as to whether it is time to end concerns over the estrogenic effects of BPA, particularly since it has repeatedly been impossible to reproduce the initial positive effects (Sharpe, 2010).

The difficulties surrounding the discussions on low-dose effects of BPA are illustrated by a study performed by Schönfelder et al. (2002a). This study is often cited to support the opinion that low-dose BPA exposure alters the development of estrogen-sensitive organs in rodents (for example: Somm et al., 2009; Kum et al., 2009; Newbold et al., 2009; Vandenberg et al., 2008; Vom Saal and Welshons, 2006). Interestingly, this study has been classified as inadequate for the evaluation process by an expert panel (Chapin et al., 2008). Therefore, we intensively revisited the controversially discussed study of Schönfelder et al. (2002a) in which Sprague-Dawley dams were administered 0.1 and 50 mg/kg bw/day BPA

<sup>2</sup> See, for example, the criteria on reliability, relevance, and adequacy of data and studies by Klimisch et al., 1997.

Table 3. Examples of guideline-compliant BPA studies.

Study	Design	Result	Reference
Three-generation reproductive toxicity study in rats	<ul style="list-style-type: none"> <li>Six BPA dose groups (0.001–500 mg/kg/day; administration in diet);</li> <li>OPPTS guideline 870.03800</li> </ul>	<ul style="list-style-type: none"> <li>No effects in the low-dose range (0.001–5 mg/kg/day)</li> <li>Reduced body and organ weights at 50 mg/kg/day</li> <li>Effects on renal and hepatic histopathology: 500 mg/kg/day</li> <li>Reproductive and developmental toxicity: 500 mg/kg/day</li> <li>At doses &lt;500 mg/kg/day: <u>no</u> effects on prostate weights, acquisition of puberty</li> <li>Systemic NOAEL: 5 mg/kg/day</li> <li>Reproductive NOAEL: 50 mg/kg/day</li> </ul>	Tyl et al., 2002
Two-generation rat study	<ul style="list-style-type: none"> <li>Daily gavage doses of 0.02–200 µg/kg/day</li> <li>US EPA GLPs and OPPTS TG with added endocrine-sensitive and neurobehavioral end points</li> </ul>	<ul style="list-style-type: none"> <li>No effect at any dose (consider that this is a low-dose study with 0.2 mg/kg/day as the highest dose)</li> </ul>	Ema et al., 2001
One- and two-generation reproductive toxicity studies in mice	<ul style="list-style-type: none"> <li>Two vehicle control groups, six BPA dose groups (0.003–600 mg/kg/day; administration in diet)</li> <li>Dietary estradiol as positive control (0.2 µg/kg/day to 8 mg/kg/day)</li> <li>OECD GLPs</li> </ul>	<ul style="list-style-type: none"> <li>No effects in the low-dose range (0.003–5 mg/kg/day)</li> <li>Reduced body weights, increased renal and liver weights and further effects at 600 mg/kg/day. However, no adverse effects on adult reproductive structures or functions at 600 mg/kg/day.</li> <li>Effects on liver histopathology at 50 mg/kg/day</li> <li>Systemic NOAEL: 5 mg/kg/day</li> <li>Reproductive NOAEL: 50 mg/kg/day</li> </ul>	Tyl et al., 2008a, 2008b, 2008c
Developmental neurotoxicity study in rats	<ul style="list-style-type: none"> <li>Dietary administration of 0, 0.01, 0.1, 5, 50, and 150 mg/kg/day (mean target doses) from gestation day 0 through lactation day 21. Evaluation of F1 offspring.</li> <li>OECD test guideline 426</li> </ul>	<ul style="list-style-type: none"> <li>No treatment-related neurobehavioral effects</li> <li>No evidence of neuropathology and no effects on brain morphometry</li> <li>NOAEL for systemic toxicity derived from maternal and offspring body weight reductions: 5.85 and 13.1 mg/kg/day (calculated) during gestation and lactation, respectively</li> </ul>	Stump et al., 2010

from gestation days 6 to 21 (Schönfelder et al., 2002a). The authors report that they observed “striking morphological changes” in the vagina of postpubertal offspring” and “that the full-length ER $\alpha$  is not expressed during estrus in the vagina of female offspring...” The “thickness of the total epithelium was reduced...” following exposure to 0.1 mg/kg BPA (Figure 1B in Schönfelder et al., 2002a) when compared to the control group (Figure 1A in Schönfelder et al., 2002a). The 50 mg/kg dose caused a similar effect (as the 0.1 mg/kg dose), but this was less pronounced (Figure 1D in Schönfelder et al., 2002a). Taking these results very seriously, we, however, have to report that severe criticism has been expressed (Chapin et al., 2008) arguing that the study of Schönfelder et al. (2002a) does not give any information on the numbers of animals used and the number of offspring examined. Also, it lacks any statistical analysis of the results. Even the number of litters represented was not stated. It is important that the litter and not the individual rodent pup be used for statistical analysis, because there may be large differences between individual litters (Goodman et al., 2006; Willhite et al., 2008). It is a critical difference if the “six offspring in the 50 mg/kg per day BPA group” (Schönfelder et al., 2002a) are pups from six separate litters rather than from only one litter. This has not been specified by Schönfelder

et al. (2002a). The study was criticized by the EFSA (2006) because the results (Schönfelder et al., 2002a) were taken from experiments where none of the control and BPA treatments were performed at the same time. Therefore, the EFSA expert panel came to the conclusion that this casts doubt on the robustness of the observations and makes this study unsuitable for risk assessment (EFSA, 2006). After our own thorough assessment of this study we came to the same conclusions as the expert panel mentioned above (Chapin et al., 2008). This means that according to the rigorous requirements in the regulatory process, we have to conclude that the data of this study do not meet the criteria necessary for a regulatory safety assessment.

Because of the high public interest into the question whether low-dose BPA may cause adverse effects and based on quality issues with some studies claiming important adverse effects, the National Institute of Environmental Health Sciences (NIEHS) has just announced an investment of approximately 30 million US dollars into BPA-related research (Spivey, 2009), including 10 two-year studies on the potential contribution of low-dose BPA to obesity, diabetes, reproductive disorders, asthma, sexually dimorphic behaviors, cardiovascular diseases, as well as prostate, breast, and

uterine cancer. Although BPA is already one of the most intensively studied chemical compounds, the new wave of studies will offer the chance to achieve an even clearer picture and to provide the basis for a final decision on the existing controversy. As part of the “lessons learnt,” we come to the conclusion that in all future studies, as a minimum requirement, the following, principally self-evident, rules should be followed: (i) study design, endpoints, and statistical procedures must be defined in a study plan before the onset of the study; (ii) sufficient numbers of doses should be tested (studies with only one or two doses are not sufficient) and the number of animals per group, accounting also for litter effects, must be sufficient according to statistical power analyses; (iii) validated or clearly interpreted endpoints should be tested; (iv) the route of exposure should be known for the exposure routes in humans; (v) study plan and the obtained raw data should be transparent and made available to the scientific community—Taken into consideration the regulatory background, and the fact that no intellectual property needs to be protected, it is not acceptable if raw data are not made available to other scientists or competent authorities for reanalysis; and (vi) each study must be performed in a way such that it can be reproduced by others. The use of commercially available laboratory animals is preferred. If, however, in-house strains are used, they should be made available to scientists wishing to reproduce the study.

### **How can differences between industry-sponsored and publicly sponsored studies be explained?**

There is strong controversy on the weight given to studies that were performed under GLP and according to standard Organisation for Economic Co-operation and Development (OECD) protocols (mostly industry sponsored due to the requirement for GLP and guideline studies for regulatory submission) and exploratory studies (mostly sponsored by public funding). Some scientists (e.g., Myers et al., 2009) claim that “funding by chemical corporations accounts for most other studies that conclude low doses of BPA are safe,” whereas the majority of publicly funded studies show that low BPA doses cause harm (<http://endocrinedisruptors.missouri.edu/vom-saal/vom-saal.html>) and that studies funded by industry “used insensitive, out-of-date protocols and assays that are incapable of finding many of the adverse effects.” They conclude that these studies are flawed (Myers et al., 2009). However, this way of interpreting differences between guideline-compliant and exploratory research, mainly performed at universities and research institutes with public funding, is naive. It ignores the basically different conditions, goals, and strategies of both types of research. Exploratory research at universities is often performed to discover new mechanisms or to analyze how a compound interferes with mechanisms that represent cutting-edge topics of current academic research.

Animal numbers usually need to be low, as technical conditions and resources do not allow the treatment, observation, and analysis of high numbers of animals. Statistical evaluation of explorative studies is usually performed by tests accepting differences between compound exposed samples and controls as significant if a  $p$  value smaller than .05 is obtained without applying an appropriate adjustment for multiple testing. Hence, the findings can only be interpreted as exploratory results and need at least one further study confirming the findings. A  $p$  value of .05 means that for 100 endpoints in a study where a compound has no influence, 5 endpoints result in (false-)positive results. Considering that many more than 1000 explorative studies, with each study examining multiple endpoints, have been published, we have to expect a relatively high number of “false-positive” findings. Importantly, they are “false positive” only for statistical reasons, not because of false claims or flaws. It is also important to consider that it is much easier to publish positive than negative results of explorative studies in high-ranking journals. This leads to the well-known phenomenon of publication bias. Therefore, it is not helpful to count how many academic studies are positive versus negative and to decide by majority vote whether a health hazard has to be expected or not. An important aspect is also that explorative studies may identify a chemical-induced biological event, but this event may not translate into an adverse health effect. The long-term low-dose safety studies on BPA demonstrate this.

The conditions and the purpose of guideline-compliant regulatory studies are completely different from those of academic studies (Tyl, 2009a, 2009b). Guideline-compliant toxicity testing aims to identify specific and validated endpoints associated with adverse health effects in a well-characterized animal model. The study design and tests to be performed must be documented prior to the study. High numbers of animals and adequate statistical procedures have to be applied. Guideline-compliant toxicity studies also determine a wide range of parameters (hematology, clinical chemistry, etc.) and include full histopathological assessment of all organs considered relevant. Usually, a battery of such guideline-compliant and quality-controlled tests is used in risk assessment in order to analyze a sufficient number of different endpoints. Under conditions of guideline-compliant tests, the probability of false-positive results is much lower, particularly because of the higher number of animals, a priori defined study design, and highly sophisticated statistics.

Usually, there is a fruitful interplay between explorative (university) studies and guideline-compliant (industry or contract research organization) studies. In principle, it is possible that novel effects are detected in explorative studies that are not captured by guideline-compliant studies. If such observations are made, the reproducibility and relevance with respect to adverse health effects should be investigated. If confirmed, observations from explorative studies should indeed be used for risk assessment.



There are techniques initially used in explorative studies that were later validated and included into toxicity testing guidelines. An example is the use of primary hepatocytes for drug metabolism studies that, after initial use in explorative studies, has subsequently been approved by the United States Food and Drug Administration (US FDA) (Hewitt et al., 2007). Considering the different condition and purpose as well as the fruitful interactions, it is not adequate to play off explorative against guideline-compliant studies just because the percentages of positive results differ between both study types.

## Toxicokinetics

### Humans

BPA is well absorbed by the oral route. Völkel et al. reported urinary recovery in human volunteers of 97% of the dose in males and 84% in females after oral administration (2002, 2005), indicating extensive absorption of orally administered BPA (key findings summarized in Tables 4 and 5). It should be underscored that Völkel et al. (2002, 2005, 2008) performed their studies by dosing deuterated BPA in order to differentiate between BPA present due to contamination and BPA resulting from dosing. Recently, the study by Völkel has been criticized by Vandenberg et al. (2010a, 2010b) who deemed it to be of limited validity. Therefore, we compiled all critical comments of these authors and assessed their relevance (Table 6). We came to the conclusion that the criticism by Vandenberg et al. (2010a, 2010b) is not justified and that the pharmacokinetic studies by Völkel et al. (2002, 2005), the results of which are consistent with those of other toxicokinetics studies with BPA in nonhuman primates (Table 5), are useful for describing BPA pharmacokinetics in humans.

BPA is metabolized to its glucuronide and sulfate conjugates (Hanioka et al., 2008; Kim et al., 2003; Ye

et al., 2005). In humans, glucuronidation was described to be catalyzed by the uridine 5'-diphospho (UDP)-glucuronosyltransferase UGT2B15 (Hanioka et al., 2008). More recent work was unable to determine whether UGT2B7 or UGT2B15 is the relevant enzyme (Mazur et al., 2010) but showed that intestinal metabolism does not play an important role in humans. Sulfation is mediated most probably by the sulfotransferase isoform SULT1A1, as SULT1A1 preferentially conjugates phenols (Campbell et al., 1987a, 1987b). In addition, among an array of bacterially expressed SULT isoforms, SULT1A1 had the highest  $k_{cat}/K_M$  value for BPA conjugation, indicating that this SULT isoform has a relevant contribution to the conjugation of BPA in vivo (Nishiyama et al., 2002). In individuals unintentionally exposed to BPA, glucuronides account for 85% and sulfates for 15% of the oral dose in urine (Ye et al., 2005), whereas sulfates were not identified as metabolites of BPA in experimentally exposed humans (Völkel et al., 2002). Similarly, in a Korean study, the average excretion of the glucuronide in adult men and women was determined to be 80% and 40%, respectively, whereas the proportion of sulfate ester excretion was higher in women than in men (about 40% and 20%, respectively) (Kim et al., 2003). From in vitro data Kurebayashi et al. (2010) calculated that 92% of hepatic clearance is due to glucuronidation and 8% due to sulfation. In studies with experimental oral exposure of 5 mg deuterated BPA, no parent compound but BPA-glucuronide was quantified in plasma and urine. In some biomonitoring urine samples, only small amounts of unchanged BPA were found the amount ranging between a few percent up to 9.5% of the total amount recovered in urine (Dekant and Völkel, 2008; Völkel et al., 2008; Ye et al., 2005). In the study of Völkel et al. (2008), using the identical analytical procedure and equipment as for biomonitoring samples, no deuterated BPA was detected in the urine of a male human

Table 4. Key conclusions from the toxicokinetics study of Völkel et al. (2002) in human subjects using deuterated BPA [d(16)-BPA].

- 5 mg of d(16)-BPA (60–80 µg/kg/bw) were orally administered to human volunteers; blood and urine sampling was performed at predetermined time points.
- Maximum blood concentrations were measured approximately 80 minutes after administration.
- The half-life of d(16)-BPA was less than 6 hours.
- The administered doses were completely recovered in urine as d(16)-BPA-glucuronide.

Note. Similar data were obtained by Doerge et al. (2010b), Kurebayashi et al. (2002), and Tominaga et al. (2006) in non-human primates.

Table 5. Kinetic parameters of bisphenol A in blood at comparable single oral doses in primates and rodents.

Species	Dose (µg/kg bw)	$C_{max}$ (nmol/L)	$t_{max}$ (h)	$t_{1/2}$ (h)	AUC (nmol × h × L <sup>-1</sup> )	Reference
Rhesus monkeys	100	590 (BPA conjugates) 0.84 (free BPA)	0.5 (total BPA)	3.5 (total BPA) 0.39 (free BPA)	920 (BPA total) 1.5 (free BPA)	Doerge et al., 2010b
Cynomolgus monkeys	100	456 (total BPA)	1.0 (total BPB)	9.6 (total BPA)	1162 (BPA-glucuronide)	Kurebayashi et al., 2002
Human subjects	60–80	820 (BPA conjugates) <10 (free BPA)	1.35 (total BPA)	5.3 (terminal) 1.5 (initial)	2792 (BPA-glucuronide)*	Völkel et al., 2002
Sprague-Dawley rats, female	100	73 (total BPA) 0.39 (free BPA)	2 (both free and total)	4.6 (total BPA) 3.0 (free BPA)	680 (total BPA) 2.6 (free BPA)	Doerge et al., 2010a

\*Calculated based on a clearance of 7.8 L/h and an oral bioavailability of 99%.

Table 6. Data on the fate of BPA in humans have been published by Völkel et al., 2002, 2005.\*

Criticism	Assessment of the commission
1. "Several inconsistencies are present in this report and therefore raise questions about its reliability (Völkel et al., 2002). For instance, the authors report two different values when maximal plasma concentrations ( $C_{max}$ ) were achieved (1.35 hours versus 4 hours)."	The authors definitely do not report two different time points for maximal plasma concentrations. In fact, they wrote, "The results from this study show that maximal plasma concentrations of d <sub>16</sub> -bisphenol A glucuronide were reached approximately 80 min after oral administration" (Völkel et al., 2002; p. 1285). The blood concentrations of d(16)-BPA-glucuronide are summarized in Figure 6 (Völkel et al., 2002; p. 1285) and show only one peak approximately 80 minutes after administration of BPA and not two peaks as claimed by Vandenberg et al. (2010a, 2010b). As clearly stated in the publication, two separate studies were performed giving identical doses. The first study was intended to provide mass balance/recovery and observe blood levels over a longer time period. Urinary and blood concentrations were determined after oral administration of BPA. In this first experiment, blood concentrations were measured at 4, 8, 12, 16, 24, and 32 hours after BPA administration and the highest concentration of BPA-glucuronide was observed after the earliest examined period of 4 hours (Figure 5B in Völkel et al., 2002). In the second study blood levels of BPA-glucuronide were analyzed in shorter intervals of 0, 51, 81, 111, 141, 201, 261, 321, and 381 minutes (Figure 6 in Völkel et al., 2002) to determine initial kinetics. This resulted in a kinetic profile with a single peak after approximately 80 minutes. In fact, extrapolation of the half-life from the first study exactly predicts the levels at the sampling points in the second study and blood concentrations of BPA-glucuronide at overlapping sampling points from the two studies are essentially identical. It should be considered that the first experiment was needed to assess recovery and kinetics in blood at later time points, which cannot be studied with the design required for assessment of distribution and initial elimination from blood, because the latter requires short sampling intervals and thus blood sampling by a venous port. It should also be noted that the authors (Völkel et al., 2002) never mentioned "two different values for the time when $C_{max}$ was achieved" in their paper. In conclusion, this criticism of Vandenberg et al. (2010a, 2010b) is not justified.
2. "Additionally, the BPA-glucuronide levels reported in blood are higher than the total BPA concentrations measured in the same individuals."	The blood concentrations of d(16)-BPA-glucuronide and total d(16)-BPA after glucuronidase treatment are very similar with overlapping error bars as shown in Figure 6 (Völkel et al., 2002). There is no significant difference. Vandenberg et al. may have misinterpreted Figure 6 because the peak values of d(16)-BPA-glucuronide determined by LC-MS/MS are slightly above total d(16)-BPA determined after glucuronidase treatment. However, the difference is in the range of the experimental error. In conclusion, Figure 6 of Völkel et al. (2002) clearly shows that concentrations of total and glucuronidated BPA in blood are very similar after oral administration of BPA.
3. "Finally, the authors indicate that they measured BPA metabolism in 3 women, a group of 3 men, and then in a separate group of 4 men, yet the groups of male volunteers clearly overlap, making the data compiled from combining these two groups questionable."	The characteristics (gender, age, height, body weight) of the individuals who joined the pharmacokinetic study are given in Table 1 (Völkel et al., 2002, p. 1282), together with identification numbers. Urinary excretion was analyzed in individuals A, B, C, E, F, and G, as described in Materials and Methods. First, since a range-finding study was not carried out for all male individuals, data from four subjects had to be handled separately. Second, BPA-glucuronide concentrations in blood of these individuals were followed over relatively long periods (4, 8, 12, 16, 24, and 32 hours) after administration of BPA (Figure 5) in the first experiment. In the second experiment, the kinetics in blood were determined using shorter intervals (Figure 6). This second experiment was performed with individuals G, M, N, and O (Figure 6). Therefore, only one individual (G) took part in both studies and the determined blood concentrations are highly consistent between the two studies (see above). The commission sees no reason why this constellation should compromise the results, particularly since the study design was clearly described.
4. Vandenberg et al. (2010a, 2010b) criticized that "there was a lack of acknowledgement of the likelihood of different toxicokinetics when BPA exposure is continuous compared to a single administration. Ginsberg and Rice suggested that results from the Völkel et al. study were more consistent with delayed excretion from long-term internal storage or cycling between conjugation and deconjugation (Ginsberg and Rice, 2009)."	First, it should be emphasized that the aim of the study of Völkel et al. (2002) was to analyze toxicokinetics of BPA after administration of a single dose. Therefore, this argument cannot be used to criticize the study. Second, basic pharmacokinetic calculations permit the prediction of blood concentrations after repeated doses once the half-life and clearance of a compound are known. The relevant parameters for such calculations are available from Völkel et al. (2002). It should also be kept in mind that the argument of Ginsberg and Rice (2009) is highly questionable, since it is based only on a higher blood concentration at only one time point (24 hours) in one gender where error bars clearly overlap and the differences are not statistically significant. In addition, none of the available other primate studies give any evidence for cycling of BPA. Long-term internal storage or cycling is also very unlikely, since most of the administered compound is recovered in the urine within 24 hours (Völkel et al., 2002).

Table 6. continued on next page

Table 6. Continued.

Criticism	Assessment of the commission
5. Vandenberget al. also criticized that "the potential for BPA to have actions at low levels (in the ng/ml range) was not considered" in the Völkel study.	Again, this is not the aim of a toxicokinetics study. The commission has the impression that Vandenberget al. (2010a, 2010b) confuse the goals of a toxicokinetics study and risk assessment. We comment on this under "criticism 12." The relevance of low doses of BPA has been discussed in section "Do oral low BPA doses below 5 mg/kg bw/day cause adverse health effects in laboratory animals?" of this article. It is important to note that recent pharmacokinetic modeling studies have shown that the highest reported oral exposures for BPA in the general population would result in blood concentrations much lower than concentrations claimed to cause some effects in vitro (Mielke and Gundert-Remy, 2009).
6. "Third, this toxicokinetics study was designed to assess the metabolism of BPA following oral exposure because until very recently (Stahlhut et al., 2009), it was assumed that most if not all BPA exposure in humans was occurring via the oral route. All sources of BPA have not been identified, so non-oral exposures cannot be discounted."	This is not a valid argument against the quality of the study of Völkel et al. (2002) because this study aimed at analyzing the pharmacokinetics after oral administration of BPA. We have discussed the question of oral versus non-oral exposure in section "Specific exposure conditions" of this article. Briefly, it should be kept in mind that BPA is quantitatively excreted via the urine. Techniques to estimate overall uptake of BPA from urinary concentrations are generally accepted (and discussed in Mielke and Gundert-Remy, 2009).
7. "Finally, the possibility of differences in toxicokinetics under different physiologic paradigms was overlooked.... The differences between sexes and age groups in urinary levels of BPA found in biomonitoring studies from CDC (Calafat et al., 2005, 2008) raise the possibility that toxicokinetics of chemicals and drugs including BPA are likely to be different in fetuses and neonates compared to adults."	Analysis of differences between neonates and adults, etc., was not the aim of the study of Völkel et al. (2002). Again, Vandenberget al. (2010a, 2010b) mix up the goals of a toxicokinetics study and the risk assessment process. For risk assessment, safety factors are used to consider possible differences between neonates and adults. In this context, the finding that a recently published PBPK model predicted the internal exposure in newborns to be approximately 3 times higher than in adults should be considered (Mielke and Gundert-Remy, 2009). The prediction model took into account conjugation by sulfotransferases, which is already active in newborns (Milke and Gundert-Remy, 2009). Moreover, toxicokinetic data from neonatal non-human primates are available (Doerge et al., 2010b). The 2008 Calafat study simply measured urine from neonates in intensive care units and found that urine samples mostly contained conjugates, and the two groups of neonates investigated in this study had major differences in average exposures. The authors suggest that the neonates were exposed to BPA via intravenous medical devices used at one of the sites. The study of Calafat did not examine toxicokinetics of BPA.
8. "BPA kinetics were examined in six individuals administered BPA, although no information was provided about the characteristics of these subjects making it difficult to draw any conclusions from this study (Völkel et al., 2005)."	The authors give detailed characteristics of the subjects participating in the study. In Table 1 (Völkel et al., 2005, page 1749), gender, age, height, and body weight of all six individuals are stated. Further information is given in the Materials and Methods section: "All subjects enlisted in the study had to refrain from alcoholic beverages and medicinal drugs 2 days before and throughout the experiment. Subjects did not abuse alcohol and were nonsmokers. Subjects were healthy, as judged by medical examination and clinical blood chemistry." Therefore, the statement by Vandenberget al. (2010a, 2010b) that "no information was provided" is unsubstantiated.
9. "The authors suggested that there were no differences in kinetics between volunteers, yet a closer look shows a wide variation in BPA measurements between individuals (Völkel et al., 2005). In the three men examined, 85 % of the administered BPA dose was recovered in urine after 5 h, mostly as BPA-glucuronide. In the three women examined, 75 % of BPA was recovered as BPA-glucuronide after the same period of time."	There is no "wide variation in BPA measurements between individuals." The data (Völkel et al., 2002) show a $93 \pm 19$ nmol recovery for men and $83 \pm 16$ for women. Using a second technique analyzing total BPA after enzymatic cleavage of glucuronides, recovery was $106 \pm 16$ nmol and $92 \pm 16$ nmol, for men and women, respectively. Therefore, the ranges overlap and there is thus no evidence for a wide variation between individuals.

Table 6. continued on next page

Table 6. Continued.

<p>10. "In two of six individuals, unconjugated BPA was detected in the urine at levels of approximately 1 ng/ml. This finding directly contradicts the conclusions reached by the study authors, who suggested that 100 % first-pass metabolism would promptly convert BPA to its conjugated metabolites."</p>	<p>There is certainly no contradiction and the authors have carefully considered this aspect (Völkel et al., 2002, 2005, 2008). When deuterated d(16)-BPA was used, Völkel et al. did not detect free BPA in the urine. By contrast, when studies were performed with non-deuterated BPA, very low levels of free BPA were detected in some samples. In a further study, the authors even administered both d(16)-BPA and non-deuterated BPA (Völkel et al., 2008). Interestingly, no free d(16)-BPA but free non-deuterated BPA was detected in urine. The authors discussed contamination as a possible reason for the observation that only non-deuterated free BPA was observed. It should also be considered that 1 ng/ml of free BPA represents a very low concentration. A concentration of 1 ng/ml corresponds to a daily intake of approximately 0.025 µg/kg bw, which is 2000-fold below the tolerable daily intake (TDI) of 50 µg/kg bw/day. It should be taken into account that glucuronides of BPA in urine may release free BPA. Therefore, the presence of free BPA in urine is difficult to interpret, particularly if only concentrations close to the LOD are detected.</p>
<p>11. Vandenberg et al. (2010a, 2010b) criticized the fact that LODs of Völkel et al. (2002) were higher than in other studies.</p>	<p>It should be kept in mind that the study (Völkel et al., 2002) is a toxicokinetics study and not a biomonitoring study. In a toxicokinetic study, sensitivity should be sufficient to study the time course of a controlled dose of a chemical for a sufficient period of time. In the case of the study by Völkel et al. (2002), concentrations of d(16)-BPA in urine were above the LOD even 42 hours after administration. The LOD of 10 nM d(16)-BPA in blood is also sufficient, considering that the peak concentrations of the BPA-glucuronide in blood were approximately 800 nM and concentrations could be determined over the time frame covered by the study design. In conclusion, the commission does not see any reason why the toxicokinetics study of Völkel et al. (2002) is compromised because of the sensitivity of the analytical procedure.</p>
<p>12. Vandenberg et al. (2010a, 2010b) criticized the conclusion of Völkel et al. (2002) concluded from their study "that there is no risk from current human exposure levels (Völkel et al., 2002)."</p>	<p>No such statement can be found in the article of Völkel et al. (2002). Moreover, it was designed as a toxicokinetics study and therefore does not make any conclusion on risk. Again, Vandenberg et al. confuse the goals of a pharmacokinetics study and of risk assessment. Toxicokinetics studies may have important implications for risk assessment; however, risk assessment is not based on toxicokinetics studies per se. Risk assessment is based on NOAELs or benchmark doses from toxicity studies in animals and on exposure assessment. Kinetics can only be used to justify acceptable margins of exposure. It should also be kept in mind that the EFSA has never based its assessment on the outcome of any of the mentioned BPA kinetics studies but used the species differences in kinetics to conclude that the margin of exposure of 100 is conservative.</p>

\*Aspects of these studies have been criticized (Vandenberg et al., 2010a, 2010b). Here, we assess the relevance of the criticism of Vandenberg et al. (2010a, 2010b).



subject who ingested deuterated BPA (dose, 60 ng/kg bw), whereas trace amounts of non-deuterated BPA were found in these urine samples. The authors attributed this finding to BPA contamination of the urine, e.g., by house dust containing BPA. Others (Doerge et al., 2010a, 2010b; Cao et al., 2010; Sajiki et al., 1999) have also discussed the problem of contamination. In addition to contamination, free BPA detected in urine may be formed from urinary BPA-glucuronide that has undergone enzymatic hydrolysis by autologous or bacterial  $\beta$ -glucuronidase in the urinary bladder (Helander and Dahl, 2005; Ho and Ho, 1985; Paigen and Peterson, 1978; Zenser et al., 1999). Waechter et al. (2007) pointed out that BPA-glucuronide may be unstable in urinary samples under some conditions during storage or analytical work-up steps. Hence, there is overwhelming evidence that unchanged BPA is excreted in the urine in only very low quantities and confounding by artefacts cannot be excluded (Twaddle et al., 2010).

The half-life of BPA can be estimated from urinary excretion data, assuming that the rate-limiting step is metabolic transformation and not urinary excretion of the conjugated metabolites. From the data of Tsukioka et al. (2004), a half-life of 1.5 hours can be derived in humans. A similar half-life, namely 2.28 hours, has been calculated by Shin et al. (2004) in their physiologically based biokinetic (PBBK) model, whereas Cho et al. (2002) calculated an even shorter half-life of 0.73 hours. Using PBBK modeling, Shin et al. (2004) calculated a volume of distribution at steady state of 1.94 L/kg bw and Cho et al. (2002) calculated it to be 1.71 L/kg bw, with corresponding clearances of 26.6 ml/min/kg bw and 29.0 ml/min/kg bw in humans. Using their clearance value, Shin et al. (2004) calculated that a serum concentration of BPA of 1.49 ng/ml (measured by Takeuchi and Tsutsumi, 2002) corresponds to a daily dose of 100 mg BPA, which is more than 2 orders of magnitude higher than the highest exposure of 0.9  $\mu$ g/kg/day taken from biomonitoring data of Calafat et al. (Calafat et al., 2005, 2008). Hence, the measured concentrations of Takeuchi and Tsutsumi (2002) using an unreliable enzyme-linked immunosorbent assay (ELISA) (see below in "How can biomonitoring support risk evaluation?") are highly implausible.

#### **Non-human primates**

Following intravenous (i.v.) administration of BPA ( $^{13}\text{C}_{12}$ -BPA stable isotope-labeled substance to avoid background contamination) to adult monkeys, rapid elimination with a half-life of 3.6 hours was observed (Doerge et al., 2010b). Five minutes after administration, more than 70% of circulating BPA was conjugated, suggesting a rapid metabolism. In contrast to rats (Doerge et al., 2010a), no enterohepatic recirculation was observed in monkeys. After oral administration (100  $\mu$ g BPA/kg bw), absorption of BPA was nearly complete (Doerge et al., 2010b). The concentrations of free BPA in serum of adult monkeys were very low (<1 nM) and absolute bioavailability of BPA, based on the relation of the areas under the plasma concentration-time curve

( $\text{AUC}_{\text{oral}}/\text{AUC}_{\text{i.v.}}$ ), was 0.2%, indicating a high first-pass effect. The mean serum concentration-time profile for total BPA in rhesus monkeys administered an oral dose of 100  $\mu$ g/kg bw was similar to that of human volunteers administered a dose of 50–90  $\mu$ g/kg bw BPA (Völkel et al., 2002). The pharmacokinetic parameters of Doerge et al. (2010) are in fair agreement with those previously reported for aglycone and conjugated BPA in adult male and female cynomolgus monkeys (Kurebayashi et al., 2002; Tominaga et al., 2006). Similar results were recently reported by Taylor et al. (2010) in monkeys, confirming the findings of the other authors.

#### **Rats**

The most recent study using stable isotope-labeled substance showed a half-life of 0.66 hours following i.v. administration. In this species, more than 50% of circulating BPA was already conjugated at the earliest analyzed time point of 5 minutes, demonstrating a high metabolic turnover. The plasma concentration-time profiles exhibited a second peak in the concentration of total BPA, which points to an enterohepatic recirculation after biliary excretion, as has been previously described by other authors (Kurebayashi et al., 2003; Upmeier et al., 2000; Pottenger et al., 2000). The kinetic parameters (AUC, elimination half-time, clearance, and volume of distribution) reported by Doerge et al. (2010a) in female rats were comparable to the results of Yoo et al. (2000, 2001) for male Sprague-Dawley rats. A high first-pass effect can be assumed because peak concentrations of total BPA after oral administration contained much lower percentages of unchanged parent compound than observed after i.v. injections (Doerge et al., 2010a). The absolute oral bioavailability of BPA was reported to be 2.8%. This low bioavailability is similar to that reported by Yoo et al. (2001) of 5.3% in adult male Sprague-Dawley rats.

#### **Mice**

Taylor et al. (2010) published data on aglycone and conjugated BPA in female CD-1 mice demonstrating linear kinetics over a broad range of doses (2  $\mu$ g/kg to 100,000  $\mu$ g/kg), a short half-life (~4 hours), and no accumulation after repeated dosing. In the first serum sample (0.5 hours after dosing) aglycone BPA was ~1% of the total (aglycone plus conjugated BPA), indicating a high first pass. As in rats, the plasma concentration-time profiles exhibited a second peak in the concentration of total BPA, which points to an enterohepatic recirculation.

### **Importance of the exposure route**

Exposure of the general population to BPA occurs mostly via food and beverages that have been in contact with polycarbonate plastic. As oral BPA undergoes extensive presystemic elimination whereby glucuronidation accounts to more than 90% (see above), the activity of the metabolites is important to know for risk assessment. As shown by several authors, BPA-mono-glucuronide is

no longer active as an estrogen (Matthews et al., 2001; Snyder et al., 2000). Since most of human BPA exposure of humans occurs via ingestion (EU, 2003, 2008; Geens et al., 2010; Wilson et al., 2007), laboratory animal studies using the oral route are the most relevant for human risk assessment.

Many of the studies showing adverse effects at low doses of BPA used subcutaneous injections. In others BPA was injected into discrete regions or delivered by osmotic pumps. Unless blood and/or tissue concentrations are monitored to compare to systemic/internal BPA concentrations in humans, the results of such studies are not appropriate for risk assessment purposes. This has usually not been carried out in studies using injections or after implantation of pumps.

Plausible explanations for the effects observed following non-oral administration of BPA are the lack of first-pass metabolism and the slow release of BPA from the oil suspensions injected. Since the administration route in animal tests for human risk assessment should be the same route as human exposure, we see no reasonable argument why administration routes other than oral should be tested. The exception might be exposure by the dermal route. Deviation from testing animals by the oral route is only justified if laboratory animals show a much higher first-pass detoxification than humans. However, as we have shown above in “Toxicokinetics,” this is not the case for BPA.

### **Can rodents be used to extrapolate to the human situation with respect to estrogenic activity?**

The current TDI for BPA is based on NOAELs derived from studies using rats and mice. Therefore, it is important to know if these rodent species are similarly susceptible to BPA as humans or—critically—whether humans are much more sensitive. An important aspect to consider in this context is the endogenous production of the estrogen 17 $\beta$ -estradiol (E2) in diverse species. A comparison of plasma levels of E2 (taking into account different phases of the reproductive cycle) across mammalian species revealed that mouse, rat, and dog regulate their normal cycle at comparatively low levels of estrogen, whereas the estrogen levels in monkey and human during particular phases of the cycle are 1 to 2 orders of magnitude higher (Günzel et al., 1989). Accordingly, it is plausible that higher exposures to exogenous estrogens will be required to provoke changes in the endocrine regulation of the human organism compared to certain animal species (if other factors such as the pharmacokinetics are similar—see below). Furthermore, across species, there is a pronounced variability in the number of estrogen receptors, even in the same organ; and the affinity of a certain xenobiotic to these receptors may also vary. Therefore, Günzel et al. (1989) concluded that even when there are effects that are clearly mediated via hormone receptors, a direct, quantitative extrapolation from experimental animals to humans is not justified.

A second critical question is whether there are major pharmacodynamic interspecies differences in susceptibility to estrogens. Available data support the conclusion that rats exhibit a similar sensitivity to EE2 compared to humans, or are slightly more sensitive. In addition, the question of possible insensitive rat strains has been intensively addressed (Gray et al., 2010; Health Canada, 2008; Chapin et al., 2008; NTP, 2008). Several experts came to the conclusion that no single rat strain is highly sensitive or resistant to estrogens. Finally, it should be considered that in current drug development, the pharmacological activity of new hormonal drug candidates is still characterized successfully in rodent species before entering into clinical studies in humans. To our knowledge, results of such experiments have not revealed any “low-dose phenomena” as they have been claimed to occur with BPA.

The misunderstanding of the “estrogen-insensitive rat” came from a letter (vom Saal, 2010) asserting that doses of EE2 (of less than 0.5  $\mu\text{g}/\text{kg bw}/\text{day}$ ) included in oral contraceptives did not cause effects in the rat study of Ryan et al. (2010a). It should be noted, however, that such comparisons on the basis of dose alone are misleading. There are notable differences in the oral bioavailability and, hence, systemic availability of EE2 in rats (approximately 3%) and humans (approximately 45%; Kuhnz et al., 1999). Accordingly, a comparison of sensitivity should incorporate a correction factor accounting for differences in the systemic exposure (i.e., area under the curve [AUC] of plasma concentration over time) as a basis, rather than simply a comparison of the external dose based on body weight; it should also refer to the same endpoint of pharmacological activity. This leads to the critical question of interspecies differences in BPA pharmacokinetics. It is well known that humans and monkeys excrete the BPA-glucuronide predominantly via the urine (Kurebayashi et al., 2002; Völkel et al., 2002). In contrast, rats excrete BPA-glucuronide predominantly via the bile into the feces, resulting in enterohepatic circulation (Inoue et al., 2001; Kurebayashi et al., 2003; Upmeier et al., 2000). In addition, the glucuronidation rate of BPA is higher in liver microsomes obtained from rats compared to humans (Elsby et al., 2001a, 2001b). Considering these interspecies differences—enterohepatic circulation in rodents but not in primates and higher glucuronidation rates for rats compared to humans—interspecies extrapolation from the rodent to the primate or human situation is complex. An overview of interspecies differences in BPA kinetics is given in Table 5. The AUC for BPA-glucuronide after single oral doses of BPA seems to be higher in humans compared to cynomolgus and rhesus monkeys as well as rats. On the other hand, higher exposures to exogenous estrogens may be required to provoke changes in the endocrine regulation in humans or monkeys compared to rats, because rats regulate their normal cycle at lower levels of estrogen, as explained above (Günzel et al., 1989). Therefore, it can be expected that the extrapolation factor of 10 for interspecies differences to obtain the current TDI of 50  $\mu\text{g}/\text{kg bw}/\text{day}$  is conservative.

## Are there susceptible subpopulations?

The risk assessment of a chemical includes consideration of susceptible subpopulations, which require a specific risk assessment. Risk is determined by hazard and by exposure to the chemical. The hazard is expressed in quantitative terms by the NOAEL, which is adjusted by an assessment factor for interspecies differences and intraspecies/intersubject variability. The generally used default factor to account for the intraspecies variability is 10, which is subdivided into a factor of 3.3 for toxicokinetic and an additional factor of 3.3 for toxicodynamic variability (WHO, 2005). A factor higher than 10 may be necessary to cover a higher variability given in special subpopulations. This can be due to particular toxicokinetic features (mainly because of lower metabolism and/or excretion), or due to particular toxicodynamic features (concentration-response relationship shifted to a lower concentration range at which effects are elicited). Lower metabolism and/or excretion, as well as a shifted concentration-response relationship, may be present in a specific subpopulation and cause concern even if the subpopulation is exposed at the same exposure level as the general population. Conversely, a subpopulation with normal toxicokinetic and toxicodynamic patterns may be at risk because their exposure is higher than the worst-case exposure scenario calculated for the general population.

The following section discusses the toxicokinetics and toxicodynamics of BPA with the aim of evaluating whether there may be defined subpopulations at risk. Exposure considerations are also discussed (see “How can biomonitoring support risk evaluation?”), with the exception of the special situation in neonates in intensive care units and the situation in newborns and babies fed using polycarbonate bottles.

### Toxicokinetics in children

One type of subpopulation at higher risk than the “normal” population is defined by the feature that at the same external exposure their internal body burden, expressed as concentration in blood/plasma, is higher than the internal body burden of the “normal” population. A higher internal body burden might be due to increased absorption or decreased elimination, both of which would lead to an increase in AUC. Because absorption of BPA is nearly 100%, increased absorption due to factors such as age or disease is not a consideration for BPA. Lower metabolic activity would be the key underlying cause for a decrease in elimination as BPA undergoes extensive conjugation via glucuronidation and sulfation. In newborns and infants up to 6 months, glucuronidation activity is known to be reduced, whereas older children have similar activities to adults (Allegaert et al., 2008; Edginton et al., 2006; Gow et al., 2001; Miyagi and Collier, 2007; Zaya et al., 2006). Hanioka et al. (2008) demonstrated that UGT2B15 is one of the enzymes responsible for glucuronidation of BPA in microsomes from adult humans. Experimental

data on UGT2B15 in human development have not been reported so far. However, data are available for UGT2B7, which belongs to the same UGT2B subfamily. The data indicate that the glucuronidation activity of UGT2B7 is 5% of the adult level in newborns, increasing to 30% after 3 months, 80% after 6 months, and 100% at the age of 1 year. This information can be used with some confidence to describe the age-dependent pattern of UGT2B15. It is conceivable that at a given external exposure the internal body burden is higher in children (up to 12 months) compared to adults because of reduced metabolic capacity. BPA plasma concentrations in newborns and infants were predicted by two groups using age-specific toxicokinetic models, which implemented the lower metabolic activity via glucuronidation. Using a model with elimination by glucuronidation as the only pathway, Edginton and Ritter (2009) simulated plasma concentrations in the newborn, who at a given external exposure were 11-fold higher compared to concentrations in adults. They, however, did not take into consideration that SULT1A1 mediates sulfation of BPA and that it is already expressed at high levels, even in intrauterine life. Likewise, SULT1E1 and SULT2A1, which are also capable of BPA sulfation, have also been detected and investigated in fetal tissues (Coughtrie, 2002; Gamage et al., 2006; Pacifici and Marchi, 1993; Duanmu et al., 2006; Miki et al., 2002; Stanley et al., 2005). In a second modeling approach, Mielke and Gundert-Remy (2009) implemented both metabolic pathways—glucuronidation (85% of the excretion in adults) and sulfation (15% excretion in adults). They modified glucuronidation in an age-dependent manner, with 5% of the adult value for glucuronidation in the newborn. In the adult, they predicted similar plasma concentrations to those measured by Edginton and Ritter (2009), whereas the predicted plasma concentrations were only 3-fold higher in newborns than in adults and 1.6-fold higher in 3-month-olds than in adults. The result is due to the fact that in subjects with reduced glucuronidation, a greater proportion of BPA is metabolized to the sulfate metabolite. However, as SULT1A1 has a lower intrinsic metabolic clearance compared with UGT2B15, the concentration of BPA is increased (Kurebayashi et al., 2010). Hence, by definition, newborns and infants up to 3 months would qualify as a subpopulation at risk. In addition to the lower glucuronidation activity, it has to be taken into consideration that SULT1A1 is polymorphically expressed in humans. Dependent on the ethnicity, the prevalences of the wild-type allele \*1 and the less active alleles \*2 and \*3 were reported to be 50–90%, 10–30% and 0–3%, the latter being active only in African Americans in a prevalence of up to 20% (Coughtrie 2002). The functional consequences of this known polymorphism towards the metabolism of BPA are uncertain at this time, as 15% enzyme activity versus 50% enzyme activity in blood platelets have been reported versus no impaired functionality in the recombinant enzyme at all (Coughtrie, 2002). For adults, these observations are of no great quantitative importance, as only 15% of the



BPA elimination is via this pathway. For newborns and infants, however, where sulfation might be an important pathway, according to the modeling results of Mielke and Gundert-Remy (2009), a reduced enzyme activity would have consequences on the blood concentration of free BPA. If the polymorphism is functionally relevant for BPA metabolism, then the model without the sulfation pathway used by Edginton and Ritter (2009) would describe a worst-case scenario, namely a totally non-functional sulfation pathway. To which extent this scenario is realistic remains open.

In conclusion, in the special subpopulation of newborns and babies up to 6 months, metabolism is impaired and intraspecies variability is greater than the default factor of 3.3. Two considerations play a role for the pharmacokinetic intraspecies variability: first, the fact that the value of 5% glucuronidation activity in newborns describes the median value, which means that there might be newborns and specifically premature infants with a lower than 5% glucuronidation activity. Second, there is the possibility of an impaired sulfation pathway in newborns homozygous for *SULT1A1\*2* or *SULT1A\*3*, but only in a small fraction of the population. From these facts, it follows that the default factor of 3.3, which is used to account for the toxicokinetic variability in the general population, seems to be large enough to cover the variability in the newborn population.

#### **Toxicokinetics in other groups**

Pregnant women have been stated to have a generally impaired metabolism of xenobiotics. This is not true; moreover, and specifically for glucuronidation, there are data showing slightly elevated activity compared to non-pregnant women (Anderson, 2005; Hodge and Tracy, 2007). Often, pregnant women are referred to as being at risk, whereas it is meant that the embryo/fetus would be exposed and at a specific risk. When assessing the risk of in utero exposure, the exposure of the fetus depends on maternal blood concentrations. Maternal metabolism is the mechanism by which most xenobiotics, including BPA, are eliminated. Because "accumulation" of BPA in the fetus does not occur and the human placenta does not metabolize BPA, only a very limited amount of the compound gains access to the fetus, which has been shown by ex vivo perfusion of the human placenta (Balakrishnan et al., 2010). As *SULT1A1* activity is present from the 26th week of life, fetal metabolism contributes to a certain extent to the overall elimination of BPA (see below) (Pacifi and Marchi, 1993; Duanmu et al., 2006). The elderly have also been stated to exhibit slower metabolism, which is true only to a limited extent concerning phase 1 reactions (Butler and Beck, 2008; He et al., 2006) but not for phase 2 reactions (Court, 2010).

In conclusion, the fetus is not at risk during the prenatal phase, because it is protected by the maternal metabolism. The relevance of BPA exposure via baby bottles for this subpopulation is discussed below. There

is no indication that the elderly or pregnant women are at risk, since their metabolic capacity is not impaired.

#### **Tissue deconjugation of BPA-glucuronide and BPA-sulfate**

Ginsberg and Rice (2009) opened up the discussion that tissue BPA concentrations might be higher than calculated due to deconjugation of BPA-glucuronide and BPA-sulfate in tissues. Although there is no doubt on the presence of the deconjugation enzymes  $\beta$ -glucuronidase and sulfatase in several tissues, it should be emphasized that for risk assessment, quantification of reactions and the chemical species present in equilibrium are indispensable. Recently, experimental data on deconjugation of BPA-glucuronide have been published, which allows quantification of this process in the rat fetus (Nishikawa et al., 2010). According to this publication, uterine (maternal) exposure to 113 nmol BPA-glucuronide resulted in a fetal exposure of 147 pmol BPA-equivalents (109.26 pmol BPA-glucuronide in the fetus plus 31.35 pmol BPA in the amniotic fluid plus 6.45 pmol BPA in the fetus), corresponding to 0.13% of the given dose. In the further quantification, we assume that BPA is present in the amniotic fluid because it is excreted by the fetal kidneys as BPA-glucuronide and then converted back to BPA. This assumption leads to the conclusion that 6.45 pmol BPA (in the fetus) are formed from of 147 pmol BPA-glucuronide, which accounts for 4.4% of the dose passed through the placental membrane. Since 0.13% of BPA-glucuronide passes the placental membrane and from this, 4.4% is converted back to BPA in the fetus, it can be estimated that only 0.006% of the maternal BPA-glucuronide is converted back to BPA in the fetus. Edginton and Ritter (2009) simulated an average BPA-glucuronide concentration of 0.15  $\mu\text{g/L}$  (375 pM) for a realistic worst-case external exposure of 1  $\mu\text{g/kg bw/day}$  in a human adult. Using the rat data of 0.006% conversion in the fetus, this would mean that due to the exposure with BPA-glucuronide, the fetal BPA concentration would be 0.0225 pM. Balakrishnan et al. (2010) reported that in human placenta perfusion experiments, BPA does cross the placenta. The concentration at the fetal side is 0.9-fold the concentration at the maternal side. Hence, for a dose of 1  $\mu\text{g/kg bw/day}$  and a resulting concentration of 0.003  $\mu\text{g/L}$  (Edginton and Ritter, 2009; Mielke and Gundert-Remy, 2009), fetal BPA exposure via blood is 0.0027  $\mu\text{g/L}$  ( $0.9 \times 0.003 \mu\text{g/L}$ ) (11.8 pM). The BPA concentration is added to the BPA concentration formed by the deglucuronidation of BPA-glucuronide, giving a value of 0.0225 pM at 1  $\mu\text{g/kg bw/day}$  (see above). This estimate reveals that at a maternal exposure of 1  $\mu\text{g/kg bw/day}$  (which represents a highly conservative estimate; see below in "How can biomonitoring support risk evaluation?"), fetal BPA exposure is 11.8 pM, whereby passage of BPA-glucuronide through the placenta and deconjugation contributes to the exposure with an amount of 0.2%.

Deconjugation of estrone sulfate by steroid sulfatase (STS) is an important mechanism for the intracellular



availability of estrone. Estrogen sulfatase is a microsomal enzyme and is ubiquitously distributed in several mammalian tissues, among which the liver, placenta, and endocrine tissues exhibit relatively high activity (Iwamori, 2005). Tan and Pang (2001) characterized the process in liver cells in vitro by reporting  $K_M$  and  $V_{max}$  values. Valle et al. (2006) found that the specific enzymatic activity of STS in adipocytes was 118 pmol/10<sup>6</sup> cells per hour, approximately 50–100 times lower than in the placenta. According to Stowell et al. (2006), BPA-sulfate (BPAS) and -disulfate are substrates for STS. In their in vitro system exposing MCF-7 cells to BPAS, desulfation BPA-disulfate and uptake of BPA were observed. Stowell et al. (2006) concluded that sulfation may increase the estrogenic potential of xenobiotics. They observed increased levels of BPA in their cellular system after incubation with BPA-sulfate, because of intracellular deconjugation to the active form. However, given the fact that sulfation is a minor pathway in infants older than 1 year and adults, with only 15% of a BPA dose being sulfated, even a rapid and complete cellular uptake and deconjugation by STS would increase the available amount by 15%, which is not a dramatic increase. In newborns and infants up to 3 months, conjugation to BPA-sulfate becomes an important metabolic pathway because of the lower glucuronidation activity. Hence, deconjugation by STS might have a relevant impact on the availability of BPA at the cellular level. Data in humans on expression and activity of STS are not available. However, the maturation of sulfatase activity has been investigated in developing rats by using triiodothyronine sulfate ( $T_3S$ ) as a substrate (Huang et al., 1996). In hepatic microsomal preparations from fetal rats, desulfation activity was extremely low. There was a non-significant trend of increasing desulfation activity in rats after birth until 1 month of age. Desulfating activity increased between the 1- and 2-month-old groups to reach adult levels at the end of the second month, mainly due to increased enzyme capacity. If the results in the rats are applicable to humans, it could be concluded that due to lacking STS during the first months of life, deconjugation of BPA-sulfate does not occur to a significant extent. Hence, even in the age groups in which conjugation to BPA-sulfate becomes an important metabolic pathway, availability of BPA at the cellular level is not increased due to low expression of STS in this age group.

### Toxicodynamics

Due to its estrogenic property, BPA is expected to have an impact on physiological processes that are influenced by estrogens. To obtain an insight into the possible impact of BPA, it is helpful to compare BPA levels in maternal blood with their levels of estrogens during pregnancy. Increasing gestagen and estrogen concentrations are observed in the course of pregnancy. Whereas 17 $\beta$ -estradiol serum levels in women in the reproductive age vary between 50 pg/ml (menstruation) and 200 pg/ml (follicular development) (0.18 nM and 0.73 nM), they are 3000 pg/ml (11 nM) at 12 weeks and increase to 25,000

pg/ml (92 nM) at week 40 of pregnancy (Salas et al., 2006). When the concentrations of estradiol are compared with the predicted concentration of BPA of 0.003  $\mu$ g/L (11 pM) at the highest exposure levels in adults (1  $\mu$ g/kg bw/day), the ratio between estradiol and BPA increases from 60-fold (persistent follicle) to 8000-fold at week 40. Taking into account the much lower estrogenic potency of BPA, it is obvious that BPA does not contribute to a biologically relevant extent to the total estrogen exposure during pregnancy.

## Specific exposure conditions

### Neonatal intensive care unit

Specific exposure conditions to BPA were reported by Calafat et al. (2009) in patients of a neonatal intensive care unit. The mean urinary concentration of BPA-glucuronide in a single urine sample of this population was 30.3  $\mu$ g/L, with the highest individual measured value was 946  $\mu$ g/L. Unfortunately, no clinical details on the neonates were reported, so a number of assumptions have to be made for further calculations. Taking 300 ml as the urine volume per day and a neonate body weight of 3 kg (International Commission on Radiological Protection, 2002), this gives a median intake of 3.03  $\mu$ g/kg (maximum intake of 94.6  $\mu$ g/kg) and a median BPA steady-state plasma concentration of 0.026 ng/ml (maximum steady state concentration 0.83 ng/ml). An intake of 94.6  $\mu$ g/kg exceeds the TDI of 50  $\mu$ g/kg bw/day derived for the adult based on oral rat data. Hence, neonates in intensive care units may have a specifically high exposure to BPA, most probably because of intravenous exposure to products containing polycarbonates. Exposure on a neonatal intensive care unit is not for the whole life and this has to be taken into consideration for risk assessment. However, 20% of the calculated concentrations range above 1 nM (= 0.23 ng/ml). In in vitro studies with human adipocytes, this concentration has been reported to stimulate mouse  $\beta$ -cell insulin production and secretion by activation of the extracellular signal-related protein kinase 1/2 pathway and to inhibit adiponectin release (Vom Saal and Myers, 2008). In conclusion, neonates in intensive care units may be exposed to BPA by the intravenous route in high amounts. Under those conditions, calculated internal BPA concentrations are in the concentration range that elicits effects in in vitro studies.

### Polycarbonate (PC) bottles

There are several studies demonstrating that BPA migrates from PC feeding bottles. Leaching varies among products and experimental conditions such as temperature and duration of the procedure (Brede et al., 2003; Cao and Corriveau, 2008; EFSA, 2006). Migration can increase with repeated use of the bottles because of cleaning procedures. An upper value of migration of 50  $\mu$ g BPA/L was identified in the EU (2003), whereas in two migration studies conducted under realistic conditions of use, the highest level of BPA migration in used PC bottles was

measured to be 22 µg/L (Tan and Mustafa, 2003; Brede et al., 2003). Using the worst-case estimate concentration of 50 µg BPA/L from the EU risk assessment report (infant formulae in used bottles), EFSA (EFSA, 2006) calculated an exposure of 11 µg/kg bw/d BPA for a 3-month infant who was fed with infant formula with a PC bottle. The estimate is conservative and represents a realistic worst-case scenario. Infants up to 3 months belong to a sub-population at higher internal exposure according to their altered toxicokinetics. However, the exposure level of 11 µg/kg bw/day does not exceed the TDI modified by an additional factor of 3 to account for interindividual differences in toxicokinetics (TDI of 50 µg/kg bw/day divided by 3 = 17 µg/kg bw/day). Hence, although the exposure in the age group of 3-month-old infants fed with infant formula with polycarbonate bottles is enhanced compared to breast-fed infants, it does not raise concern.

## How can biomonitoring support risk evaluation?

### Results and interpretation of biomonitoring studies with BPA

Biomonitoring is a direct approach to estimate human exposures to chemicals from environmental and occupational sources. Due to highly sensitive analytical chemistry, biomonitoring has developed into a valuable tool in exposure assessment (Angerer et al., 2007; Boogaard, 2007; Calafat and Needham, 2007, 2009; Needham et al., 2007). However, transforming biomonitoring data to a daily dose requires a detailed understanding of the toxicokinetics of an agent. Moreover, due to the high sensitivity of modern analytical chemistry, detailed quality control and reduction of potential contamination with the analyte from other sources is needed (Calafat and Needham, 2007; Dekant and Völkel, 2008; Hoppin et al., 2006).

Regarding BPA, a large number of biomonitoring studies are available (for overviews see Dekant and Völkel, 2008; Vandenberg et al., 2007, 2010a, 2010b). Concentrations of BPA present in both urine and plasma of the general population are often close to the limits of quantification (LOQs), even using highly sensitive methods, and this, together with the many confounding factors associated with these methods, has raised specific issues in data generation and evaluation that need to be addressed (Calafat and Needham, 2009; Dekant and Völkel, 2008; Markham et al., 2010; Ye et al., 2007):

- Stability of BPA and BPA conjugates in the sample matrix and during sample processing.
- Sample handling and sample work up; for example, avoidance of polycarbonate-containing plastic tools such as tubes or syringes.
- Measures to reliably reduce background of BPA in sample blanks to levels below the LOD.
- Variability of background present in blanks.

- Use of appropriate internal standards, especially stable isotope-labeled derivatives.
- Description of sensitivity and specificity of the applied analytical method.
- Detailed quality control of the analytical method and sample processing including recovery, precision, and reproducibility.

The results of the many biomonitoring studies on BPA have recently been reviewed and will not be reiterated in detail here (Dekant and Völkel, 2008; EFSA, 2006; US FDA Memorandum, 2009a, 2009b; Vandenberg et al., 2010a, 2010b). Moreover, since only urine or blood concentrations of BPA and its metabolites are useful in exposure assessment, the comments concentrate on these two matrices.

In the large number of urine samples analyzed for BPA (>10,000), most reported concentrations of total BPA were well below 10 µg/L (Bushnik et al., 2010; Calafat et al., 2005, 2008; Lakind and Naiman, 2008, 2010); higher concentrations were present in a very limited number of samples (Garcia-Prieto et al., 2008; Moors et al., 2007). Very high concentrations of BPA and BPA conjugates were only observed in the urine of newborns from one intensive care unit (Calafat et al., 2009) and in a study from China (Mao et al., 2004). The high urinary excretion of BPA conjugates in the newborns are likely to be related to BPA release from medical equipment (see above). Most of the BPA in urine is present in the form of conjugates when separate analysis for free BPA and BPA conjugates was performed. This observation is consistent with results from toxicokinetics studies in both humans and non-human primates with BPA after oral administration (Kurebayashi et al., 2002; Tominaga et al., 2006; Tsukioka et al., 2003, 2004; Uchida et al., 2002; Völkel et al., 2002, 2005, 2008).

Blood concentrations of “free” BPA of up to 22 µg of free and 66.48 µg of “total” BPA/L have been reported in maternal and fetal blood samples at delivery (Padmanabhan et al., 2008; Schönfelder et al., 2002b), whereas many other studies reported much lower concentrations of BPA (usually less than a few micrograms of “total” BPA/L) in blood of the general population. It has been claimed that the high concentrations reported in maternal or fetal blood at delivery suggest high exposures of the general population to BPA from unknown sources, likely through pathways where BPA is not metabolized by an intensive first pass. However, a detailed analysis of the database on reported blood concentrations of BPA, considering the strengths and weaknesses of the analytical methodologies, sampling procedures, background contamination, and biological plausibility based on the toxicokinetics, needs to be performed to draw conclusions.

A part of the database on blood, serum, or plasma concentrations of BPA is based on ELISAs (enzyme-linked immunosorbent assays). Some ELISA-based report concentrations of “free BPA” in the range of a few micrograms per liter. However, ELISA assays have been

demonstrated to widely overestimate BPA concentrations actually present and are cross-reactive with BPA-glucuronide and other constituents in blood or plasma (Lee et al., 2008). Moreover, different ELISA kits gave widely differing results with identical samples, and the BPA concentrations indicated by ELISA were inconsistent with BPA concentrations determined by instrumental analytics (Fukata et al., 2006; Tominaga et al., 2006). Therefore, ELISAs are not reliable to quantify the low concentrations of BPA present in blood samples of the general population.

The studies examining BPA in maternal and fetal blood samples also have a number of drawbacks that renders them unsuitable for an assessment of population exposures (Lee et al., 2008; Padmanabhan et al., 2008; Schönfelder et al., 2002b; Vandenberg et al., 2010a, 2010b):

- The extent of medical intervention during delivery is not reported. BPA may be released from medical devices (Calafat et al., 2009). Thus, exposure may have occurred by the intravenous route, therefore representing a specific situation (see above); the measured concentrations of BPA may therefore have no relevance for the general population.
- The publications have major problems regarding methods and reporting. Enzymatic cleavage of BPA conjugates was not carried out, thus only “free” BPA was determined.
- Despite considerable efforts to reduce BPA concentrations in blanks and contrary to statements made in the text, BPA peaks remain in blanks presented in the figures (Schönfelder et al., 2002b) and the variability of the background is not given. Moreover, a number of inconsistencies between the text and the figures regarding calibration were identified (e.g., a limit of detection [LOD] cannot be determined with BPA appearing in the blanks; the peak area of the internal standard varies widely, suggesting low reliability of the reported BPA concentrations).
- In the second paper (Padmanabhan et al., 2008), a very short solvent gradient is applied and pure methanol is used as eluting solvent during most of the separation. Due to the high eluting capacity of methanol, separation efficiency is low. BPA is also eluted from the column long after the gradient is reversed back to initial conditions. This is inconsistent with theory and practice of reversed phase liquid chromatography. Therefore, this methodology cannot be considered as reliable.

A third study (Lee et al., 2008), which reported blood levels of BPA at delivery, determined BPA after hydrolysis by high-performance liquid chromatography with postcolumn fluorescence derivatization (HPLC-FLD). Some of the results were confirmed either by gas chromatography-mass spectrometry (GC/MS) after derivatization or by liquid chromatography-mass spectrometry (LC-MS) (no

details given). No information on background induced by the derivatization for GC/MS was given (which may be expected to be high). In addition, HPLC-FLD has the disadvantage of low specificity and separation efficiency.

In blood/serum/plasma samples of the general population, most studies report lower concentrations (usually <1 µg/L) either “total” or “free” BPA; if detected, BPA concentrations were often close to the LODs of the methods applied. Many of these studies did not use specific procedures to reduce contamination and do not give BPA concentrations in blank. Studies with more elaborate quality control and low or absent BPA contamination in blanks often report BPA levels below the LODs or well below 1 µg/L. Interestingly, blank concentrations of free BPA using a sensitive LC/MS-MS system were consistently in the range of 1 ng/L of plasma, identical to the “background” exposure in human blood often cited (Twaddle et al., 2010) and studies with elaborate quality control did not detect BPA in human blood samples with an LOD of 0.3 µg/L (Ye et al., 2009).

### Biomonitoring and exposure assessment

The reliable estimate of systemic doses by biomonitoring may represent an important part of the exposure assessment if a sufficient number of samples have been analyzed. Usually, biomonitoring is more precise than indirect exposure assessments relying on assumption of food consumptions and migration from food contact material or other sources such as breathable air. However, biomonitoring has to transform concentrations measured in biological samples into a daily exposure/dose. For compounds such as BPA that are rapidly metabolized and completely eliminated in the urine, urinary concentrations are much more useful indicators of human exposure compared to blood concentrations. Concentrations of BPA in blood rapidly decline after intake due to the short half-life of “free” BPA. Any exposure assessment based on blood concentrations has to calculate back to consider infrequent food intake, which is very difficult to perform without having very detailed information.

One of the problems of biomonitoring urinary BPA is the variability between different collection periods. However, possible variations in estimates based on urinary concentrations due to urine collection intervals will be averaged due to the availability of a large data set. Concentrations of BPA and its metabolites in spot urine samples may therefore be used to calculate daily BPA exposures based on average 24-hour urine volumes. Using measured urinary concentrations of BPA and considering toxicokinetics, total daily doses of BPA well below 1 µg/kg bw have been derived (Dekant and Völkel, 2008; Miyamoto and Kotake, 2006). The daily exposure of humans to BPA, established by biomonitoring, is thus well below the daily exposure, as derived from indirect estimates of exposure based on food consumption. Such assessments are based on conservative assumptions of food intake and migration data, and, furthermore,



integrate the high end of food concentrations of the agent under study. Most regulatory agencies prefer to use such data because they rely on conservative approaches for exposure assessment. However, the US FDA, EFSA, and the Japanese government have evaluated the available biomonitoring studies and concluded that the exposure estimates based on food concentrations and migration of BPA are conservative compared to daily intakes based on biomonitoring.

When translating blood or urine concentrations from biomonitoring to daily intakes, the data should also be consistent with results obtained by other means of exposure assessment. This is the case for BPA:

- The daily dose of BPA derived by biomonitoring of urinary excretion and average daily excretion equates to similarly low mean daily intakes to exposure assessments based on food concentrations of BPA and food consumption patterns, 0.008  $\mu\text{g}/\text{kg}$  bw/day (Thomson and Grounds, 2005) or 0.002  $\mu\text{g}/\text{kg}$  bw/day (Miyakawa et al., 2004) for a 60-kg adult. Calculated daily doses of BPA based on measured concentrations in air, dust, and food were between 0.052 and 0.074  $\mu\text{g}/\text{kg}$  bw/day in preschool children (Wilson et al., 2007).
- The good agreement of daily BPA intake based on food consumption with intakes based on urinary biomonitoring further support the conclusions that food is the major exposure pathway for BPA. This is consensus among all regulatory authorities performing such exposure assessments.
- Claims that daily human exposure to BPA is much higher from unknown sources by poorly defined pathways are not substantiated. Concentrations of BPA in house dust are low. BPA is not used in cosmetics and dermal absorption of BPA is limited. The dermal contact area for potentially BPA-containing materials (e.g., credit card slips) is small, dermal penetration of BPA is limited, and the contact time of human skin with such articles is short (EU, 2003). Due to the very low volatility of BPA, exposure via inhalation can also not be considered as a relevant pathway.
- In a recent publication, it was postulated that human exposures to BPA from unknown sources are much higher than previously assumed based on the rapid elimination of “free” BPA from the blood of monkeys after oral administration (Taylor et al., 2010). The authors suggested that the background blood levels of “serum unconjugated BPA” (0.3–4 ng/ml) indicate that human exposures to BPA would have to be at least 0.5 mg/kg bw, which is orders of magnitude above those estimated by a number of regulatory agencies. However, this analysis is highly deficient. Although the authors acknowledge that a significant part of orally administered BPA is excreted with urine in primates, they do not provide any explanation of what happens to the large assumed daily doses as well as completely neglect the consistent urinary

biomonitoring data. Exposures to BPA in the range of 0.5 mg/kg bw/day should result in urine concentrations well above 10 mg/L of BPA/BPA conjugates. *Such concentrations (1000-fold higher than maximal concentrations reported) have never been reported in urine biomonitoring with samples from more than 10,000 human subjects.* Therefore, there is no valid basis for the conclusions of exposures to daily doses of BPA in the range of 0.5 mg/kg bw.

In addition to being in agreement with other exposure assessments, biomonitoring data should also be biologically plausible. The high blood concentrations of “free” BPA reported in some biomonitoring studies are inconsistent with predicted blood levels of “free” BPA even when using the high end of estimated intakes based on food concentrations nutritional habits. Very low blood concentrations of “free” BPA are predicted by PBBK modeling based on the toxicokinetics of BPA in humans and primates (Edginton and Ritter, 2009; Mielke and Gundert-Remy, 2009; Teeguarden et al., 2005). Even a simple calculation using pharmacokinetic parameters for BPA support the conclusion that high blood levels of BPA claimed in some studies are not realistic. The volume of distribution of BPA is 2 L/kg (NTP-CERHR, 2007) and the half-life in blood is around 1 hour (Kurebayashi et al., 2002; Taylor et al., 2010; Doerge et al., 2010b; Völkel et al., 2002). Therefore, a blood concentration of 10  $\mu\text{g}/\text{L}$  BPA corresponds to an intravenous dose of approximately 3 mg BPA within 1 hour before blood sampling for a 70-kg person. Adjusting to an oral uptake and considering a bioavailability of “free” BPA of 1%, the resulting intake is 300 mg BPA for a 70-kg person, which is inconsistent with all exposure estimates.

### **What are the mechanisms of action of BPA? Does the multitude of mechanisms besides estrogen receptor activation make the substance more hazardous?**

The estrogenic activity of BPA was first described in 1936 (Dodds and Lawson, 1936). BPA interacts with estrogen receptors ER $\alpha$  and ER $\beta$ , with a slightly higher affinity for ER $\beta$ . BPA has also been reported to show antiandrogenic activity at approximately 5-fold higher concentrations than those causing estrogenic activity (reviewed in Bondesson et al., 2009). Moreover, low activities have been reported for the pregnane X receptor (PXR), the estrogen receptor-related receptor (ERR), and the thyroid hormone receptor (TR) (Mnif et al., 2007; Okada et al., 2008; Abad et al., 2008; Matsushima et al., 2007; Moriyama et al., 2002; Liu et al., 2010). BPA was also reported to inhibit TH signaling but at higher concentrations than those that interact with estrogen receptors (Fini et al., 2009), and to cause increases in levels of uterine heat shock proteins (hsps), mainly hsp90 $\alpha$  and glucose-regulated protein (grp)94 (Papaconstantinou et al., 2002). The fact that BPA, in addition to its effect on estrogen receptors, also interferes with other receptors



has been used to argue that “the risk assessment of endocrine disrupting compounds, such as BPA, is hampered by large scientific uncertainties” (Bondesson et al., 2009). However, does the interaction with other receptors make the compound more hazardous? It should be considered that it is not surprising that a hormonally active chemical is not specific for a single receptor that is known also for several hormonal drugs. The potential to interact with a receptor is not per se indicative of a toxicologically relevant effect. The fact that BPA activates estrogen receptors as well as other receptors fits into a scenario frequently observed for hormonally active chemicals as well as for hormonal drugs interacting with several receptors with different affinities. In addition, it can never be excluded with certainty that further relevant toxic mechanisms have not yet been discovered. However, an important argument is that current risk assessment of BPA is based on a large number of adverse endpoints in a multitude of animal experiments (see above). These *in vivo* studies capture adverse health effects potentially induced by all receptor interactions, without requiring an *a priori* knowledge on the involved mechanisms. Of course, it is an advantage if the mechanisms responsible for observed adverse health effects are known because this offers the opportunity to refine risk evaluation, for example by comparing critical mechanisms in rodent and humans in order to identify possible interspecies differences. However, the risk to humans is only underestimated if critical toxic mechanisms are more active in humans compared to the animal species that were used for the toxicity studies. Considering the available studies on BPA, there is no evidence for such a critical (toxic) mechanism that is specific for humans or where human cells or tissues are more sensitive than the respective tissues or cells from animals.

Evaluating the relevance of ER $\alpha$ - and ER $\beta$ -mediated toxicities, it should be noted that BPA is about 10,000-fold less potent than estradiol (Gray, 2008). Because of the low levels of human exposure to BPA, it is unlikely that toxicity is mediated via estrogen receptors in humans. It should also be considered that genomic studies in rodents exposed to low doses of BPA did not result in expression of estrogen receptor-dependent genes in rat uterus or fetal rat testis (Ashby and Odum, 2004; Naciff et al., 2005, 2010). Induction of estrogen receptor-dependent genes by BPA was only observed at moderate to high dose levels with no evidence of non-linear dose-response. Evaluating the relevance of the weak estrogenicity of BPA, it should also be considered that humans are exposed to a variety of dietary natural compounds with higher estrogenic activity and at higher doses than BPA (Bolt et al., 2001; Safe 2000, 2004).

### **Epidemiological studies in the general population**

In a limited number of epidemiological studies BPA exposure data were related to health outcomes. The majority of the studies have a cross-sectional design where a single urinary BPA level is used as exposure estimate. Health outcomes were analyzed mostly by

self-reported methods (e.g., questionnaires), and had a long latency period such as for cardiovascular disease (Melzer et al., 2010) or diabetes (Lang et al., 2008; Melzer et al., 2010). The same holds true for cross-sectional studies on semen quality and sperm DNA damage (Meeker et al., 2010a, 2010b), serum testosterone, estradiol, and sex hormone-binding globulin (Galloway et al., 2010). Given the short half-life of BPA and the long latency of the health outcomes addressed, the results of the cross-sectional studies concurrently are hard to interpret. Therefore, the above-mentioned studies report associations that can at best raise hypotheses rather than demonstrate causal relationships. Also the case-control studies on breast cancer with relatively small case numbers (Yang et al., 2009) suffer from the time lag between an actual single urinary excretion of BPA as the exposure estimate and the time of occurrence of the disease. At present, there are no studies confirming the results available.

### **Recent governmental responses**

Risk management is a decision-making process to select the optimal measures for reducing a risk to an acceptable level. It follows the risk assessment step (optimally incorporating the description of uncertainties in the risk assessment process) and involves consideration of political, social, economic, and engineering factors. Risk perception by society and hence regulatory authorities also reflects the knowledge of toxicology and the culture of the society and may change with time as more information becomes available. The risk management process is iterative, taking into account any new information. Risk management decisions can therefore involve measures to prevent the process producing the risk, measures to reduce or eliminate exposures, and activities to alter perceptions or valuation. The basic options in risk management are setting toxicologically based guidance values or applying a more rigorous precautionary approach, e.g., restricting concentrations/doses to levels achievable by the best available technology.

National bans on BPA in baby bottles, which have been adopted in recent months by some countries, are based on a precautionary approach. There is no scientifically proven increased risk discernable for the age group of infants fed with polycarbonate bottles. Against the background of an ongoing controversy, it is easily understood that the current heterogeneous situation in governmental responses reflects political motivations in the respective countries rather than a scientifically justified systematic risk management. The committee therefore refrains from any scientific comment but tries to present an objective overview on the actual situation.

### **European Union**

BPA is permitted for use in food contact materials in the European Union, under Commission Directive 2002/72/EC of 6 August 2002 relating to plastic materials and

articles intended to come into contact with foodstuffs. In its risk assessment on BPA published in January 2007, the EFSA set a TDI of 0.05 mg/kg bw/day for BPA. EFSA found that intakes of BPA through food and beverages were well below the TDI, even for infants and children. EFSA's risk assessment of BPA was updated in July 2008 and October 2010 and the TDI (0.05 mg/kg bw/day) was reconfirmed (EFSA, 2008, 2010a, 2010b).

EFSA found that intakes of BPA through food and beverages were well below the TDI, even for infants and children. Typical BPA migration levels from BPA-based food contact materials are <10 µg/kg food and thus are well below the regulatory specific migration level for BPA of 600 µg/kg food (based on the TDI, assuming a person of 60 kg eating 1 kg of food per day, with an additional safety factor of 5). Using conservative migration levels, EFSA concluded in their 2007 opinion that the dietary exposure to BPA from polycarbonate plastic bottles and epoxy resin-coated food and beverage cans is well below the TDI. The updated risk assessment of June 2008, as well as EFSA's updated opinion of July 2008, states that food contact materials such as polycarbonate plastic baby bottles, drinking bottles, and epoxy resin-coated food and beverage cans are safe for their intended uses (EFSA, 2008). In its opinion, EFSA examined the safety of BPA-based food contact applications for all age groups, including fetuses and newborns.

EFSA updated its advice on BPA in September 2010. Following a review of recent human toxicity data and animal studies on the toxicity of BPA at low doses, scientists of EFSA's Panel on Food Contact Materials, Enzymes, Flavourings and Processing Aids (CEF) concluded they could not identify any new evidence that would lead them to revise the current TDI for BPA of 0.05 mg/kg bw set by EFSA in its 2006 opinion and reconfirmed in its 2008 opinion (EFSA, 2010 a, 2010b). The Panel also stated that the data currently available do not provide convincing evidence of neurobehavioral toxicity as an endpoint of concern for BPA. The present opinion follows the requests of the European Commission to the CEF Panel to evaluate the dietary developmental neurotoxicity study of BPA in rats by Stump et al. (2010) and recent scientific literature (2007 to July 2010) in terms of relevance for the risk assessment of BPA and impact on the current tolerable daily intake (TDI) of 0.05 mg BPA/kg bw/day and to provide advice on the Danish risk assessment underlying the Danish ban of BPA in food contact materials for infants aged 0–3 years.

### Denmark

On the basis of an assessment by the National Food Institute at the Technical University of Denmark (DTU Food) (<http://www.food.dtu.dk/Default.aspx?ID=8590>) released 22nd March 2010, the Danish government decided on 26th March 2010 to invoke the principle of precaution and introduce a temporary national ban on BPA in materials in contact with food for children aged

0–3 years (infant feeding bottles, feeding cups, and packaging for baby food). From 1st July 2010, BPA is not allowed in the products covered by the ban.

The overall assessment of DTU Food concluded that the new neurodevelopmental study by Stump et al. (2010) does not shed new light nor change the uncertainties about the impact of small doses of BPA on the development of the nervous system and the behavior of rodents. The conclusion from DTU Food is that this study does not give clear evidence of BPA harming the behavioral endpoints examined. However, it raises uncertainties about the impact on learning capacity. In their opinion, the study revealed reduced learning capacity of young male rats at low doses of BPA. According to the opinion of DTU Food, the finding of reduced learning capacity of newborn males may be a sign of low-dose effect of BPA. However, they also discuss that this observation may just be coincidental. The Danish Veterinary and Food Administration adopted DTU Food's opinion. On this basis, the Danish Veterinary and Food Administration concluded that the precautionary principle dictates the introduction of protective measures with respect to children aged 0–3 years until new studies document that low doses of BPA do not have an impact on development of the nervous system or on the behavior of rats.

### France

On 25th March 2010, the French Senate called on the government to suspend the commercialization of BPA-based polycarbonate baby bottles in France, until the French Food Safety Authority (AFSSA) issued conclusions on their ongoing assessments of BPA. A ban on manufacturing, importing, exporting, and selling of baby bottles made of BPA-based products has been approved by the National Assembly on 11th May 2010. The ban was endorsed under the Grenelle II sustainable development initiative. This is a temporary ban until AFSSA develops new methods for evaluating risks linked to BPA in food containers used by children under the age of 3. The French ban came into force on 30th June 2010 (<http://www.legifrance.gouv.fr/affichTexte.do;jsessionid=?cidTexte=JORFTEXT000022414734&dateTexte=&oldAction=rechJO&categorieLien=id>).

### Switzerland

The Swiss Federal Office of Public Health (FOPH) (February 2009) evaluated the scientific assessments of various international governmental agencies and came to the conclusion that exposure to BPA poses no risk for consumers including neonates and infants.

### Australia and New Zealand

Food Standards Australia New Zealand (FSANZ) has evaluated the safety of BPA and plasticizers in baby bottles and concluded that levels of intake of BPA or plasticizers are very low and do not pose a risk to infant health. In contrast, a voluntary phase out by major retailers began

on 1st July 2010. It is the result of discussions between the Australian Government and retailers.

The Australian Government appreciates there has been a level of public concern relating to BPA in baby bottles and, therefore, has worked with retailers to introduce the phase out (<http://www.foodstandards.gov.au/scienceandeducation/newsroom/mediareleases/mediareleases2010/governmentannounc-esb4822.cfm>).

### Canada

Under the Government of Canada's Chemicals Management Plan (December 2006), BPA was identified as a high-priority substance for assessment of human health risk. Environment Canada and Health Canada considered in their joint Screening Assessment from October 2008 that the neurodevelopmental and behavioral data set of BPA for rodents, though highly uncertain, is suggestive of potential effects at doses of the same order of magnitude to 1–2 orders of magnitude higher than exposures to BPA of the general population. Given that toxicokinetics and metabolism data from experimental animal and limited human studies indicate potential sensitivity to the maternal-fetal unit and infant, and that animal studies suggest a trend towards heightened susceptibility during early stages of development in rodents, it is considered appropriate to apply a precautionary approach when characterizing risk. In conclusion, BPA should be considered as a substance that may be entering the environment in a quantity or concentration or under conditions that constitute or may constitute a danger to human life or health in Canada.

A provisional TDI of 25 µg/kg bw/day was preestablished by Health Canada as a conservatively safe level for BPA presence in food and was confirmed in the 2008 Health Risk Assessment of BPA from Food Packaging Applications (Health Canada, 2009b). The prohibition of polycarbonate baby bottles that contain BPA came into force on 11th March 2010 (Canada Gazette Part II, 31st March 2010), thus prohibiting the advertisement, sale, and import of these products in Canada. Furthermore, the Canadian Government works to develop and implement codes of practice to reduce levels of BPA in infant formula as low as reasonably achievable. The ban was enforced after the market for polycarbonate baby bottles had virtually disappeared. The decision was made in spite of the results of four migration studies carried out by Health Canada and published in 2009. The studies specifically investigated migration from polycarbonate baby bottles, canned soda, canned infant formula, and bottled water, and found no or extremely low migration levels, thus confirming existing data of very low exposure (Health Canada, 2009a, 2009b, 2009c, 2009d).

### USA

The National Toxicology Program Center for the Evaluation of Risks to Human Reproduction, part of the National Institutes of Health, completed a review of

BPA in September 2008 and expressed “some concern” for effects on the brain, behavior, and prostate gland in fetuses, infants, and children at current human exposures to BPA. In the update of the draft assessment “BPA for Use in Food Contact Applications” in January 2010, the FDA shares at this interim stage the perspective of the National Toxicology Program that recent studies provide reason for some concern. The FDA also recognizes substantial uncertainties with respect to the overall interpretation of these studies and their potential implications for human health effects of BPA exposure. These uncertainties relate to issues such as the routes of exposure employed, the lack of consistency among some of the measured endpoints or results between studies, the relevance of some animal models to human health, differences in the metabolism (and detoxification) of and responses to BPA both at different ages and in different species, and limited or absent dose response information for some studies. The rating “some concern” about potential effects of BPA based on studies using novel approaches to test for subtle effects, which had been stated by the FDA/NHIES in 2008, will be addressed by a specific ongoing FDA research program. Regarding interim public health recommendations, the FDA supports reasonable steps to reduce exposure of infants to BPA in the food supply. In addition, the FDA will work with industry to support and evaluate manufacturing practices and alternative substances that could reduce exposure of the population and is supporting the industry's actions to stop producing BPA-containing bottles and infant feeding cups for the US market. The FDA is facilitating the development of alternatives to BPA for the linings of infant formula cans. But FDA still considers the TDI as valid.

### Japan

The human health risk assessment of the Japanese New Energy and Industrial Technology Development Organization (NEDO), the Research Center for Chemical Risk Management (CRM), and the National Institute of Advanced Industrial Science and Technology (AIST) from November 2007 concluded that both the human health risks and ecological risks posed by BPA were below the levels of concern. MOEs (margin of exposure) for the endpoints reduction in body weight gain, multinucleated giant hepatocytes in the liver, and reproductive toxicity were sufficiently large, even in the highest exposure group. Therefore, it will be unnecessary to prohibit or restrict the use of BPA at this time.

### Acknowledgments

We thank Prof. Wolfgang Dekant (Institute of Toxicology, University of Würzburg, Würzburg, Germany) and Prof. Regine Kahl (Institute of Toxicology, Heinrich-Heine-University, Düsseldorf, Germany) for valuable scientific discussion. The commission also thanks Ms. Susanne Lindemann for helpful bibliographic work.



## Declaration of interest

Dr. H. Schweinfurth: I am an employee of Bayer Schering Pharma, which is a division of Bayer AG. The Material Sciences division of Bayer AG is one of the producers and users of BPA. In my activities as a Nonclinical Advisor I am involved in the development of drugs for my employer, although I have no responsibilities for industrial chemicals such as BPA nor a direct relationship to the latter division.

Prof. U. Gundert-Remy: In my capacity as the former head of the department responsible for human health risk assessment of chemicals at the governmental Federal Institute for Risk Assessment (BFR) in Germany, I have been involved in the EU risk assessment of bisphenol A.

Dr. W. Völkel: I am an employee of the Bavarian Health and Food Safety Authority and responsible for the realization of biomonitoring studies and assessment of toxicological research and biomonitoring studies of chemicals such as bisphenol A. I have been involved in the following studies on BPA: Human exposure to bisphenol A by biomonitoring: Methods, results and assessment of environmental exposures (W. Dekant, W. Völkel. *Toxicol Appl Pharmacol* 2008;228:114–134). This study was supported in part by the Polycarbonate/BPA Global Group; this review represents the individual professional views of the authors and not necessarily the views of the Polycarbonate/BPA Global Group. The studies at the University of Würzburg (*Chem Res Toxicol* 2002;15:1281–1287; *Drug Metab Dispos* 2005;33:1748–1757) were supported by the German Umweltbundesamt using equipment provided by the Deutsche Forschungsgemeinschaft and the State of Bavaria. For the studies at the Bavarian Health and Food Safety Authority (*Toxicol Lett* 2008;179:155–162; *Environ Res* 2010; in press, doi:10.1016/j.envres.2010.10.001) and for the present study no external funding was obtained.

Authors Hengstler, Foth, Gebel, Kramer, Lilienblum, and Wollin report no conflicts of interest.

The authors alone are responsible for the content and writing of the paper.

## References

- Abad MC, Askari H, O'Neill J, Klinger AL, Milligan C, Lewandowski F, Springer B, Spurlino J, Rentzeperis D. (2008). Structural determination of estrogen-related receptor gamma in the presence of phenol derivative compounds. *J Steroid Biochem Mol Biol* 108:44–54.
- Akingbemi BT, Sottas CM, Koulova AI, Klinefelter GR, Hardy MP. (2004). Inhibition of testicular steroidogenesis by the xenoestrogen bisphenol A is associated with reduced pituitary luteinizing hormone secretion and decreased steroidogenic enzyme gene expression in rat Leydig cells. *Endocrinology* 145:592–603.
- Allegaert K, Vanhole C, Vermeersch S, Rayyan M, Verbesselt R, de Hoon J. (2008). Both postnatal and postmenstrual age contribute to the interindividual variability in tramadol glucuronidation in neonates. *Early Hum Dev* 84:325–330.
- Anderson GD. (2005). Pregnancy-induced changes in pharmacokinetics: A mechanistic-based approach. *Clin Pharmacokin* 44:989–1008.
- Angerer J, Ewers U, Wilhelm M. (2007). Human biomonitoring: State of the art. *Int J Hyg Environ Health* 210:201–228.
- Aschberger K, Castello P, Hoekstra E, Karakitsios S, Munn S, Pakalin S, Sarigiannis D (2010). Bisphenol A and baby bottles: Challenges and perspectives. JRC Scientific and Technical Reports, European Union.
- Ashby J, Odum J. (2004). Gene expression changes in the immature rat uterus: Effects of uterotrophic and sub-uterotrophic doses of bisphenol A. *Toxicol Sci* 82:458–467.
- Ashby J, Tinwell H, Haseman J. (1999). Lack of effects for low dose levels of bisphenol A and diethylstilbestrol on the prostate gland of CF1 mice exposed in utero. *Regul Toxicol Pharmacol* 30:156–166.
- Balakrishnan B, Henare K, Thorstensen EB, Ponnampalam AP, Mitchell MD. (2010). Transfer of bisphenol A across the human placenta. *Am J Obstet Gynecol* 202:393.e1–e7.
- Bolt HM, Janning P, Michna H, Degen GH. (2001). Comparative assessment of endocrine modulators with oestrogenic activity: I. Definition of a hygiene-based margin of safety (HBMOS) for xeno-oestrogens against the background of European developments. *Arch Toxicol* 74:649–662.
- Bondesson M, Jönsson J, Pongratz I, Olea N, Cravedi JP, Zalko D, Håkansson H, Halldin K, Di Lorenzo D, Behl C, Manthey D, Balaguer P, Demeneix B, Fini JB, Laudet V, Gustafsson JA. (2009). A CASCADE of effects of bisphenol A. *Reprod Toxicol* 28:563–567.
- Boogaard PJ. (2007). Human biomonitoring activities—Programmes by industry. *Int J Hyg Environ Health* 210:259–261.
- Borrell B. (2010). Toxicology: The big test for bisphenol A. *Nature* 464:1122–1124.
- Brede C, Fjeldal P, Skjevraak I, Herikstad H. (2003). Increased migration levels of bisphenol A from polycarbonate baby bottles after dishwashing, boiling and brushing. *Food Addit Contam* 20:684–689.
- Bushnik T, Haines D, Levallois P, Levesque J, Van Oostdam J, Viau C. (2010). Lead and bisphenol A concentrations in the Canadian population. Health Reports, Component of Statistics Canada (82-003-X) 21:1–12.
- Butler JM, Begg EJ. (2008) Free drug metabolic clearance in elderly people. *Clin Pharmacokinet* 47:297–321.
- Cagen SZ, Waechter JM Jr, Dimond SS, Breslin WJ, Butala JH, Jekat FW, Joiner RL, Shiotsuka RN, Veenstra GE, Harris LR. (1999). Normal reproductive organ development in CF-1 mice following prenatal exposure to bisphenol A. *Toxicol Sci* 50:36–44.
- Calafat AM, Needham LL. (2007). Factors affecting the evaluation of biomonitoring data for human exposure assessment. *Int J Androl* 30:1–5.
- Calafat AM, Needham LL. (2009). What additional factors beyond state-of-the-art analytical methods are needed for optimal generation and interpretation of biomonitoring data? *Environ Health Perspect* 117:1481–1485.
- Calafat AM, Kuklenyik Z, Reidy JA, Caudill SP, Ekong J, Needham LL. (2005). Urinary concentrations of bisphenol A and 4-nonylphenol in a human reference population. *Environ Health Perspect* 113:391–395.
- Calafat AM, Ye X, Wong LY, Reidy JA, Needham LL. (2008). Exposure of the U.S. population to bisphenol A and 4-tertiary-octylphenol: 2003–2004. *Environ Health Perspect* 116:39–44.
- Calafat AM, Weuve J, Ye X, Jia LT, Hu H, Ringer S, Huttner K, Hauser R. (2009). Exposure to bisphenol A and other phenols in neonatal intensive care unit premature infants. *Environ Health Perspect* 117:639–644.
- Campbell NR, Van Loon JA, Sundaram RS, Ames MM, Hansch C, Weinshilboum R. (1987a). Human and rat liver phenol sulfotransferase: Structure-activity relationships for phenolic substrates. *Mol Pharmacol* 32:813–819.
- Campbell NR, Van Loon JA, Weinshilboum RM. (1987b). Human liver phenol sulfotransferase: Assay conditions, biochemical properties and partial purification of isozymes of the thermostable form. *Biochem Pharmacol* 36:1435–1446.
- Canada Gazette. Part II, March 31, 2010.



- Cao XL, Corriveau J. (2008). Migration of bisphenol A from polycarbonate baby and water bottles into water under severe conditions. *J Agric Food Chem* 56:6378–6381.
- Cao XL, Corriveau J, Popovic S, Coughlan MC, Chepelev N, Willmore W, Schrader T, Jin X. (2010). Background bisphenol A in experimental materials and its implication to low-dose in vitro study. *Chemosphere* 81:817–820.
- Chapin RE, Adams J, Boekelheide K, Gray LE Jr, Hayward SW, Lees PS, McIntyre BS, Portier KM, Schnorr TM, Selevan SG, Vandenberg JG, Woskie SR. (2008). NTP-CERHR expert panel report on the reproductive and developmental toxicity of bisphenol A. *Birth Defects Res B Dev Reprod Toxicol* 83:157–395.
- Cho CY, Shin BS, Jung JH, Kim DH, Lee KC, Han SY, Kim HS, Lee BM, Yoo SD. (2002). Pharmacokinetic scaling of Bisphenol A by species-invariant time methods. *Xenobiotica* 32:925–934.
- Coughtrie MWH. (2002). Sulfation through the looking glass—Recent advances in sulfotransferase research for the curious. *Pharmacogenom J* 2:297–308.
- Court MH. (2010). Interindividual variability in hepatic drug glucuronidation: Studies into the role of age, sex, enzyme inducers, and genetic polymorphism using the human liver bank as a model system. *Drug Metab Rev* 42:202–217.
- Danish Ministry of Food, Agriculture and Fisheries (2010). Danish ban on bisphenol A in materials in contact with food for children aged 0–3. Available at: <http://www.fvm.dk/Default.aspx?ID=18488&PID=169747&NewsID=6014>. March 2010.
- DeKant W, Völkel W. (2008). Human exposure to bisphenol A by biomonitoring: Methods, results and assessment of environmental exposures. *Toxicol Appl Pharmacol* 228:114–134.
- Dodds EC, Lawson W. (1936). Synthetic oestrogenic agents without the phenanthrene nucleus. *Nature* 137:996.
- Doerge DR, Twaddle NC, Vanlandingham M, Fisher JW. (2010a). Pharmacokinetics of bisphenol A in neonatal and adult Sprague-Dawley rats. *Toxicol Appl Pharmacol* 247:158–165.
- Doerge DR, Twaddle NC, Woodling KA, Fisher JW. (2010b). Pharmacokinetics of bisphenol a in neonatal and adult rhesus monkeys. *Toxicol Appl Pharmacol* 248:1–11.
- Duanmu Z, Weckle A, Koukouritaki SB, Hines RN, Falany JL, Falany CN, Kocarek TA, Runge-Morris M. (2006). Developmental expression of ary, estrogen, and hydroxysteroid sulfotransferases in pre- and postnatal human liver. *J Pharmacol Exp Ther* 316:1310–1317.
- Edginton AN, Ritter L. (2009). Predicting plasma concentrations of bisphenol A in children younger than 2 years of age after typical feeding schedules, using a physiologically based toxicokinetic model. *Environ Health Perspect* 117:645–652.
- Edginton AN, Schmitt W, Voith B, Willmann S. (2006). A mechanistic approach for the scaling of clearance in children. *Clin Pharmacokinet* 45:683–704.
- EFSA (European Food Safety Authority). (2006). Opinion of the Scientific Panel on Food Additives, Flavourings, Processing Aids and Materials in Contact with Food (AFC) related to 2,2-bis(4hydroxyphenyl)propane. *EFSA J* 428:1–75.
- EFSA (European Food Safety Authority). (2007). Opinion of the Scientific Panel on food additives, flavourings, processing aids and materials in contact with food (AFC) related to 2,2-BIS(4-HYDROXYPHENYL)PROPANE. January 2007. Available at: <http://www.efsa.europa.eu/en/scdocs/scdoc/428.htm>.
- EFSA (European Food Safety Authority). (2008). Toxicokinetics of Bisphenol A—Scientific Opinion of the Panel on Food additives, Flavourings, Processing aids and Materials in Contact with Food (AFC). Available at: <http://www.efsa.europa.eu/en/scdocs/scdoc/759.htm>. July 2008.
- EFSA (European Food Safety Authority). (2010a). Scientific Opinion on Bisphenol A: Evaluation of a study investigating its neurodevelopmental toxicity, review of recent scientific literature on its toxicity and advice on the Danish risk assessment of Bisphenol A of the EFSA Panel on Food Contact Materials, Enzymes, Flavourings and Processing Aids (CEF) on request from the European Commission, Questions No. EFSA-Q-2009-00864, EFSA-Q-2010-01023 and EFSA-Q-2010-00709, adopted on 23rd September 2010. *EFSA J* 8:1829, 1–116. Available at: <http://www.efsa.europa.eu/en/scdocs/scdoc/1829.htm>. (September 2010).
- EFSA (European Food Safety Authority). (2010b). Statistical re-analysis of the Biel maze data of the Stump et al (2010) study: “Developmental neurotoxicity study of dietary bisphenol A in Sprague-Dawley rats”. Question No. EFSA-Q-2010-01142. Adopted: 30 September 2010. *EFSA J* 8:1836, 1–67. Available at: <http://www.efsa.europa.eu/fr/efsajournal/pub/1836.htm>. September 2010.
- Elsby R, Maggs JL, Ashby J, Park BK. (2001a). Comparison of the modulatory effects of human and rat liver microsomal metabolism on the estrogenicity of bisphenol A: Implications for extrapolation to humans. *J Pharmacol Exp Ther* 297:103–113.
- Elsby R, Maggs JL, Ashby J, Paton D, Sumpter JP, Park BK. (2001b). Assessment of the effects of metabolism on the estrogenic activity of xenoestrogens: A two-stage approach coupling human liver microsomes and a yeast estrogenicity assay. *J Pharmacol Exp Ther* 296:329–337.
- Ema M, Fujii S, Furukawa M, Kiguchi M, Ikka T, Harazono A. (2001). Rat two-generation reproductive toxicity study of bisphenol A. *Reprod Toxicol* 15:505–523.
- EU. (2003). European Union Risk Assessment Report. Bisphenol A, CAS No: 80-05-7. Institute for Health and Consumer Protection, European Chemicals Bureau, European Commission Joint Research Centre, 3rd Priority List. Luxembourg: Office for Official Publications of the European Communities.
- EU. (2008). European Union Risk Assessment Report. Bisphenol A, CAS No: 80-05-7, Addendum 2008. Institute for Health and Consumer Protection, European Chemicals Bureau, European Commission Joint Research Centre, 3rd Priority List, Luxembourg: Office for Official Publications of the European Communities.
- Federal Office of Public Health (FOPH). (2009). Factsheet Bisphenol A (in German). Available at: <http://www.bag.admin.ch/themen/lebensmittel/04861/06170/index.html>. (February 2009).
- Fini JB, Dolo L, Cravedi JP, Demeneix B, Zalko D. (2009). Metabolism of the endocrine disruptor BPA by *Xenopus laevis* tadpoles. *Ann N Y Acad Sci* 1163:394–397.
- Fukata H, Miyagawa H, Yamazaki N, Mori C. (2006). Comparison of ELISA- and LC-MS-based methodologies for the exposure assessment of bisphenol A. *Toxicol Mech Methods* 16:427–430.
- Galloway T, Cipelli R, Guralnick J, Ferrucci L, Bandinelli S, Corsi AM, Money C, McCormack P, Melzer D. (2010). Daily Bisphenol A excretion and associations with sex hormone concentrations: Results from the InCHIANTI Adult Population Study. *Environ Health Perspect* 118:1603–1608.
- Gamage N, Barnett A, Hempel N, Dugleby RG, Windmill KF, Martin JL, McManus ME. (2006). Human sulfotransferases and their role in chemical metabolism. *Toxicol Sci* 90:5–22.
- García-Prieto A, Lunar ML, Rubio S, Pérez-Bendito D. (2008). Determination of urinary bisphenol A by coextractive microextraction and liquid chromatography-fluorescence detection. *Anal Chim Acta* 630:19–27.
- Geens T, Apelbaum TZ, Goeyens L, Neels H, Covaci A. (2010). Intake of bisphenol A from canned beverages and foods on the Belgian market. *Food Addit Contam Part A Chem Anal Control Expo Risk Assess* 27:1627–1637.
- Gies A, Heinzow B, Dieter HH, Heindel J. (2009). Bisphenol A workshop of the German Federal Environment Agency—March 30–31, 2009 work group report: Public health issues of bisphenol A. *Int J Hyg Environ Health* 212:693–696.
- Ginsberg G, Rice DC. (2009). Does rapid metabolism ensure negligible risk from bisphenol A? *Environ Health Perspect* 117:1639–1643.
- Goodman JE, McConnell EE, Sipes IG, Witorsch RJ, Slayton TM, Yu CJ, Lewis AS, Rhomberg LR. (2006). An updated weight of the evidence evaluation of reproductive and developmental effects of low doses of bisphenol A. *Crit Rev Toxicol* 36:387–457.
- Goodman JE, Witorsch RJ, McConnell EE, Sipes IG, Slayton TM, Yu CJ, Franz AM, Rhomberg LR. (2009). Weight-of-evidence evaluation of reproductive and developmental effects of low doses of bisphenol A. *Crit Rev Toxicol* 39:1–75.

- Gow PJ, Ghabrial H, Smallwood RA, Morgan DJ, Ching MS. (2001). Neonatal hepatic drug elimination. *Pharmacol Toxicol* 88:3–15.
- Gray GM, Cohen JT, Cunha G, Hughes C, McDonnell EE, Rhombert L, Sipes IG, Mattison D. (2004). Weight of the evidence evaluation of low-dose reproductive and developmental effects of bisphenol A. *Hum Ecol Risk Assess* 10:875–921.
- Gray LE Jr. (2008). Written testimony before the Committee on Energy and Commerce, Subcommittee on Commerce, Trade, and Consumer Protection, United States House of Representatives, June 10, 2008 (unpublished document).
- Gray LE Jr, Ryan B, Hotchkiss AK, Crofton KM. (2010). Rebuttal of “Flawed experimental design reveals the need for guidelines requiring appropriate positive controls in endocrine disruption research” by Vom Saal 2010. *Toxicol Sci* 115:614–620.
- Günzel P, Putz B, Lehmann M, Hasan SH, Hümpel M, El Etreby MF. (1989). Steroid toxicology and the “pill”: Comparative aspects of experimental test systems and the human. In: Dayan AD, Paine AJ, eds. *Advances in Applied Toxicology*. London: Taylor & Francis, 19–49.
- Hanioka N, Naito T, Narimatsu S. (2008). Human UDP-glucuronosyltransferase isoforms involved in bisphenol A glucuronidation. *Chemosphere* 24:33–36.
- He P, Court MH, Greenblatt DJ, von Moltke LL. (2006). Factors influencing midazolam hydroxylation activity in human liver microsomes. *Drug Metab Dispos* 34:1198–1207.
- Health Canada. (2008). Health Risk Assessment of Bisphenol A from Food Packaging Applications. Published by Authority of the Minister of Health, Canada, August 2008. Available at: [http://www.hc-sc.gc.ca/fn-an/securit/packag-embal/bpa/bpa\\_hra-ers-eng.php](http://www.hc-sc.gc.ca/fn-an/securit/packag-embal/bpa/bpa_hra-ers-eng.php).
- Health Canada. (2009a). Survey of bisphenol A in canned drink products. Bureau of Chemical Safety, Food Directorate, Health Products and Food Branch, March 2009.
- Health Canada. (2009b). Survey of bisphenol A in bottled water products. Bureau of Chemical Safety, Food Directorate, Health Products and Food Branch. Available at: [http://www.hc-sc.gc.ca/fn-an/alt\\_formats/hpfb-dgpsa/pdf/securit/bpa\\_survey-enquete-bot-bou-eng.pdf](http://www.hc-sc.gc.ca/fn-an/alt_formats/hpfb-dgpsa/pdf/securit/bpa_survey-enquete-bot-bou-eng.pdf). (July 2009).
- Health Canada. (2009c). Survey of bisphenol A in canned powdered infant formula products. Bureau of Chemical Safety, Food Directorate, Health Products and Food Branch. Available at: [http://www.hc-sc.gc.ca/fn-an/alt\\_formats/hpfb-dgpsa/pdf/securit/bpa\\_survey-enquete-bot-bou-eng.pdf](http://www.hc-sc.gc.ca/fn-an/alt_formats/hpfb-dgpsa/pdf/securit/bpa_survey-enquete-bot-bou-eng.pdf). (July 2009).
- Health Canada. (2009d). Investigation of storage time on potential bisphenol A: Migration into canned liquid infant formula stored at room temperature. Bureau of Chemical Safety, Food Directorate, Health Products and Food Branch. (December 2009).
- Helander A, Dahl H. (2005). Urinary tract infection: A risk factor for false-negative urinary ethyl glucuronide but not ethyl sulfate in the detection of recent alcohol consumption. *Clin Chem* 51:1728–1730.
- Hengstler JG, Van der Burg B, Steinberg P, Oesch F. (1999). Interspecies differences in cancer susceptibility and toxicity. *Drug Metab Rev* 31:917–970.
- Hewitt NJ, Lechón MJ, Houston JB, Hallifax D, Brown HS, Maurel P, Kenna JG, Gustavsson L, Lohmann C, Skonberg C, Guillouzo A, Tuschl G, Li AP, LeCluyse E, Groothuis GM, Hengstler JG. (2007). Primary hepatocytes: Current understanding of the regulation of metabolic enzymes and transporter proteins, and pharmaceutical practice for the use of hepatocytes in metabolism, enzyme induction, transporter, clearance, and hepatotoxicity studies. *Drug Metab Rev* 39:159–234.
- Ho YC, Ho KJ. (1985). Differential quantitation of urinary beta-glucuronidase of human and bacterial origins. *J Urol* 134:1227–1230.
- Hodge LS, Tracy TS. (2007). Alterations in drug disposition during pregnancy: Implications for drug therapy. *Expert Opin Drug Metab Toxicol* 3:557–571.
- Hoppin JA, Ulmer R, Calafat AM, Barr DB, Baker SV, Meltzer HM, Ronnengen KS. (2006). Impact of urine preservation methods and duration of storage on measured levels of environmental contaminants. *J Expo Sci Environ Epidemiol* 16:39–48.
- Howdeshell KL, Furr J, Lambright CR, Wilson VS, Ryan BC, Gray LE Jr. (2008). Gestational and lactational exposure to ethinyl estradiol, but not bisphenol A, decreases androgen-dependent reproductive organ weights and epididymal sperm abundance in the male Long-Evans hooded rat. *Toxicol Sci* 102:371–382.
- Huang WS, Kuo SW, Chen WL, Hsieh KS, Wu SY. (1996). Maturation of hepatic desulfation activity in developing rats. *J Formos Med Assoc* 95:435–439.
- Hunt PA, Susiarjo M, Rubio C, Hassold TJ. (2009). The bisphenol A experience: A primer for the analysis of environmental effects on mammalian reproduction. *Biol Reprod* 81:807–813.
- Inoue H, Yokota H, Makino T, Yuasa A, Kato S. (2001). Bisphenol A glucuronide, a major metabolite in rat bile after liver perfusion. *Drug Metab Dispos* 29:1084–1087.
- Inoue H, Yuki G, Yokota H, Kato S. (2003). Bisphenol A glucuronidation and absorption in rat intestine. *Drug Metab Dispos* 31:140–144.
- International Commission on Radiological Protection. (2002). Basic anatomical and physiological data for use in radiological protection: Reference values. ICRP Publication 89. Amsterdam: Elsevier Science.
- Iwamori M. (2005). Estrogen sulfatase. *Methods Enzymol* 400:293–302.
- Kim YH, Kim CS, Park S, Han SY, Pyo MY, Yang M. (2003). Gender differences in the levels of bisphenol A metabolites in urine. *Biochem Biophys Res Commun* 312:441–448.
- Klimisch HJ, Andreae E, Tillmann U. (1997). A systematic approach for evaluating the quality of experimental and ecotoxicological data. *Regul Toxicol Pharmacol* 25:1–5.
- Kuhn W, Blode H, Zimmermann H. (1999). Pharmacokinetics of exogenous natural and synthetic estrogens and antiestrogens. In: Oettel M, Schillinger E, eds. *Handbook of Experimental Pharmacology*, Vol. 135/II Estrogens and Antiestrogens. Berlin, Heidelberg: Springer-Verlag, 279 (animal data), 282 (human data).
- Kum H, Kim MK, Choi HT. (2009). Degradation of endocrine disrupting chemicals by genetic transformants in *Irpex lacteus* with an inducible laccase gene of *Phlebia tremellosa*. *Biodegradation* 20:673–678.
- Kurebayashi H, Harada R, Stewart RK, Numata H, Ohno Y. (2002). Disposition of a low dose of bisphenol A in male and female cynomolgus monkeys. *Toxicol Sci* 68:32–42.
- Kurebayashi H, Betsui H, Ohno Y. (2003). Disposition of a low dose of <sup>14</sup>C-bisphenol A in male rats and its main biliary excretion as BPA glucuronide. *Toxicol Sci* 73:17–25.
- Kurebayashi H, Okudaira K, Ohno Y. (2010). Species difference of metabolic clearance of bisphenol A using cryopreserved hepatocytes from rats, monkeys and humans. *Toxicol Lett* 198:210–215.
- Lakind JS, Naiman DQ. (2008). Bisphenol A (BPA) daily intakes in the United States: Estimates from the 2003–2004 NHANES urinary BPA data. *J Expo Sci Environ Epidemiol* 18:608–615.
- Lakind JS, Naiman DQ. (2010). Daily intake of bisphenol A and potential sources of exposure: 2005–2006 National Health and Nutrition Examination Survey. *J Expo Sci Environ Epidemiol* advance online publication 17 Mar 2010; doi:10.1038/jes.2010.9
- Lang IA, Galloway TS, Scarlett A, Henley WE, Depledge M, Wallace RB, Melzer D (2008) Association of urinary bisphenol A concentration with medical disorders and laboratory abnormalities in adults. *JAMA* 300:1303–1310.
- Lee YJ, Ryu HY, Kim HK, Min CS, Lee JH, Kim E, Nam BH, Park JH, Jung JY, Jang DD, Park EY, Lee KH, Ma JY, Won HS, Im MW, Leem JH, Hong YC, Yoon HS. (2008). Maternal and fetal exposure to bisphenol A in Korea. *Reprod Toxicol* 25:413–419.
- Liu X, Matsushima A, Okada H, Shimohigashi Y. (2010). Distinction of the binding modes for human nuclear receptor ERRgamma between bisphenol A and 4-hydroxytamoxifen. *J Biochem* 148:247–254.

- Lorentzen R, Hattan D. (2010). Response to Nature Editorial. *Nature* 464:1103–1104. Available at: <http://www.nature.com/nature/journal/v464/n7292/full/4641103b.html>.
- Mao L, Sun C, Zhang H, Li Y, Wu D. (2004). Determination of environmental estrogens in human urine by high performance liquid chromatography after fluorescent derivatization with *p*-nitrobenzoyl chloride. *Anal Chim Acta* 522:241–246.
- Markham DA, Waechter JM Jr, Wimber M, Rao N, Connolly P, Chuang JC, Hentges S, Shiotsuka RN, Dimond S, Chappelle AH. (2010). Development of a method for the determination of bisphenol A at trace concentrations in human blood and urine and elucidation of factors influencing method accuracy and sensitivity. *J Anal Toxicol* 34:293–303.
- Matsushima A, Kakuta Y, Teramoto T, Koshihara T, Liu X, Okada H, Tokunaga T, Kawabata S, Kimura M, Shimohigashi Y. (2007). Structural evidence for endocrine disruptor bisphenol A binding to human nuclear receptor ERR gamma. *J Biochem* 142:517–524.
- Matthews JB, Twomey K, Zacharewski TR. (2001). In vitro and in vivo interactions of bisphenol A and its metabolite, bisphenol A glucuronide with estrogen receptors  $\alpha$  and  $\beta$ . *Chem Res Toxicol* 14:149–157.
- Mazur Ch S, Kenneke JF, Hess-Wilson JK, Lipscomb JC. (2010). Differences between human and rat intestinal and hepatic bisphenol-A glucuronidation and the influence of alamethicin on in vitro kinetic measurements. *DMD Fast Forward*. Published on August 24, 2010 at doi:10.1124/dmd.110.034819. Available at: <http://www.ncbi.nlm.nih.gov/pubmed/20736320>.
- Meeker JD, Calafat AM, Hauser R. (2010a). Urinary bisphenol A concentrations in relation to serum thyroid and reproductive hormone levels in men from an infertility clinic. *Environ Sci Technol* 44:1458–1463.
- Meeker JD, Ehrlich S, Toth TL, Wright DL, Calafat AM, Trisini AT, Ye X, Hauser R. (2010b). Semen quality and sperm DNA damage in relation to urinary bisphenol A among men from an infertility clinic. *Reprod Toxicol* 30:532–539.
- Melzer D, Rice NE, Lewis C, Henley WE, Galloway TS. (2010). Association of urinary bisphenol a concentration with heart disease: Evidence from NHANES 2003/06. *PLoS One* 5:e8673.
- Mielke H, Gundert-Remy U. (2009). Bisphenol A levels in blood depend on age and exposure. *Toxicol Lett* 190:32–40.
- Miki Y, Nakata T, Suzuki T, Darnel AD, Moriya T, Kaneko C, Hidaka K, Shiotsu Y, Kusaka H, Sasano H. (2002). Systemic distribution of steroid sulfatase and estrogen sulfotransferase in human adult and fetal tissues. *J Clin Endocrinol Metab* 87:5760–5768.
- Miyagi SJ, Collier AC. (2007). Pediatric development of glucuronidation: The ontogeny of hepatic UGT1A4. *Drug Metab Dispos* 35:1587–1592.
- Miyakawa H, Shimamura Y, Suzuki K, Ibe A, Saito K. (2004). Determination of bisphenol A in total diet study samples by GC/MS. Annual Report of the Tokyo Metropolitan Institute of Public Health, Vol. 55.
- Miyamoto K, Kotake M. (2006). Estimation of daily bisphenol A intake of Japanese individuals with emphasis on uncertainty and variability. *Environ Sci* 13:15–29.
- Mnif W, Pascucci JM, Pillon A, Escande A, Bartegi A, Nicolas JC, Cavallès V, Duchesne MJ, Balaguer P. (2007). Estrogens and antiestrogens activate hPXR. *Toxicol Lett* 170:19–29.
- Moors S, Blaszewicz M, Bolt HM, Degen GH. (2007). Simultaneous determination of daidzein, equol, genistein and bisphenol A in human urine by a fast and simple method using SPE and GC-MS. *Mol Nutr Food Res* 51:787–798.
- Moriyama K, Tagami T, Akamizu T, Usui T, Saijo M, Kanamoto N, Hataya Y, Shimatsu A, Kuzuya H, Nakao K. (2002). Thyroid hormone action is disrupted by bisphenol A as an antagonist. *J Clin Endocrinol Metab* 87:5185–5190.
- Myers JP, vom Saal FS, Akingbemi BT, Arizono K, Belcher S, Colborn T, Chahoud I, Crain DA, Farabolini F, Guillette LJ Jr, Hassold T, Ho SM, Hunt PA, Iguchi T, Jobling S, Kanno J, Laufer H, Marcus M, McLachlan JA, Nadal A, Oehlmann J, Olea N, Palanza P, Parmigiani S, Rubin BS, Schoenfelder G, Sonnenschein C, Soto AM, Talsness CE, Taylor JA, Vandenberg LN, Vandenberg JG, Vogel S, Watson CS, Welshons WV, Zoeller RT. (2009). Why public health agencies cannot depend on good laboratory practices as a criterion for selecting data: The case of bisphenol A. *Environ Health Perspect* 117:309–315.
- Naciff JM, Hess KA, Overmann GJ, Torontali SM, Carr GJ, Tiesman JP, Foertsch LM, Richardson BD, Martinez JE, Daston GP. (2005). Gene expression changes induced in the testis by transplacental exposure to high and low doses of 17 $\alpha$ -ethynyl estradiol, genistein, or bisphenol A. *Toxicol Sci* 86:396–416.
- Naciff JM, Khambatta ZS, Reichling TD, Carr GJ, Tiesman JP, Singleton DW, Khan SA, Daston GP. (2010). The genomic response of Ishikawa cells to bisphenol A exposure is dose- and time-dependent. *Toxicology* 270:137–149.
- Nagel SC, vom Saal FS, Thayer KA, Dhar MG, Boechler M, Welshons WV. (1997). Relative binding affinity-serum modified access (RBA-SMA) assay predicts the relative in vivo bioactivity of the xenoestrogens bisphenol A and octylphenol. *Environ Health Perspect* 105:70–76.
- National Food Institute at the Technical University of Denmark (DTU Food). (2010). DTU Fødevareinstituttets vurdering af industriens nye developmental neurotoxicity studie (DNT, OECD TG 426) med bisphenol A og studiets betydning for Fødevareinstituttets vurdering af bisphenol A's eventuelle skadelige effekter på udvikling af nervesystem og adfærd. 22 March 2010. Available at: <http://www.food.dtu.dk/Default.aspx?ID=8590>.
- Needham LL, Calafat AM, Barr DB. (2007). Uses and issues of biomonitoring. *Int J Hyg Environ Health* 210:229–238.
- New Energy and Industrial Technology Development Organization (NEDO), the Research Center for Chemical Risk Management (CRM), and the National Institute of Advanced Industrial Science and Technology (AIST). (2007). Bisphenol A Risk Assessment Document (AIST Risk Assessment Document Series No. 4). November 2007. Available at: [http://unit.aist.go.jp/riss/crm/mainmenu/BPA\\_Summary\\_English.pdf](http://unit.aist.go.jp/riss/crm/mainmenu/BPA_Summary_English.pdf).
- Newbold RR, Jefferson WN, Padilla-Banks E. (2009). Prenatal exposure to bisphenol A at environmentally relevant doses adversely affects the murine female reproductive tract later in life. *Environ Health Perspect* 117:879–885.
- Nishikawa M, Iwano H, Yanagisawa R, Koike N, Inoue H, Yokota H. (2010). Placental transfer of conjugated bisphenol A and subsequent reactivation in the rat fetus. *Environ Health Perspect*. 2010 Sep; 118(9):1196–203 epub 2010 Apr 9.
- Nishiyama T, Ogura K, Nakano H, Kaku T, Takahashi E, Ohkubo Y, Sekine K, Hiratsuka A, Kadota S, Watabe T. (2002). Sulfation of environmental estrogens by cytosolic sulfotransferases. *Drug Metab Pharmacokinet* 17:221–228.
- NTP (National Toxicology Program). (1982). NTP Technical Report on the carcinogenesis bioassay of bisphenol A (CAS No. 80-05-7) in F344 rats and B6C3F1 mice (feed study). Research Triangle Park, NC: NTP, NIEHS. NTP-80-35. NIH Publication No. 82-1771.
- NTP (National Toxicology Program). (1985). Teratologic evaluation of bisphenol A (CAS No. 80-05-7) administered to CD-1 mice on gestational days 6–15. Research Triangle Park, NC: NTP, NIEHS.
- NTP (National Toxicology Program). (1986). Teratologic evaluation of bisphenol A (CAS No. 80-05-7) administered to CD(R) rats on gestational days 6–15. Research Triangle Park, NC: NTP, NIEHS.
- NTP (National Toxicology Program). (2008). NTP-CERHR Monograph on the Potential Human Reproductive and Developmental Effects of Bisphenol A. NTP CERHR Monogr 2008(22):i–III. NIH publication No. 08-5994.
- NTP-CERHR. (2007). Expert Panel Report on the Reproductive and Developmental Toxicity of Bisphenol A. Research Triangle Park, NC: National Toxicology Program.
- Okada H, Tokunaga T, Liu X, Takayanagi S, Matsushima A, Shimohigashi Y. (2008). Direct evidence revealing structural elements essential for the high binding ability of bisphenol A to human



- estrogen-related receptor-gamma. *Environ Health Perspect* 116:32–38.
- Pacifici GM, Kubrich M, Giuliani L, de Vries M, Rane A. (1993). Sulphation and glucuronidation of ritodrine in human foetal and adult tissues. *Eur J Clin Pharmacol* 44:259–264.
- Padmanabhan V, Siefert K, Ransom S, Johnson T, Pinkerton J, Anderson L, Tao L, Kannan K. (2008). Maternal bisphenol-A levels at delivery: A looming problem? *J Perinatol* 28:258–263.
- Paigen K, Peterson J. (1978). Coordinacy of lysosomal enzyme excretion in human urine. *J Clin Invest* 61:751–762.
- Pottenger LH, Domoradzki JY, Markham DA, Hansen SC, Cagen SZ, Waechter JM Jr. (2000). The relative bioavailability and metabolism of bisphenol A in rats is dependent upon the route of administration. *Toxicol Sci* 54:3–18.
- Papaconstantinou AD, Fisher BR, Umbreit TH, Brown KM. (2002). Increases in mouse uterine heat shock protein levels are a sensitive and specific response to uterotrophic agents. *Environ Health Perspect* 110:1207–1212.
- Ryan BC, Hotchkiss AK, Crofton KM, Gray LE Jr. (2010a). In utero and lactational exposure to bisphenol A, in contrast to ethinyl estradiol, does not alter sexually dimorphic behavior, puberty, fertility, and anatomy of female LE rats. *Toxicol Sci* 114:133–148.
- Ryan KK, Haller AM, Sorrell JE, Woods SC, Jandacek RJ, Seeley RJ. (2010b). Perinatal exposure to bisphenol-A and the development of metabolic syndrome in CD-1 mice. *Endocrinology* 151:2603–2612.
- Safe SH. (2000). Endocrine disruptors and human health—Is there a problem? An update. *Environ Health Perspect* 108:487–493.
- Safe S. (2004). Endocrine disruptors and human health: Is there a problem? *Toxicology* 205:3–10.
- Sajiki J, Takahashi K, Yonekubo J. (1999). Sensitive method for the determination of bisphenol-A in serum using two systems of high-performance liquid chromatography. *J Chromatogr B Biomed Sci Appl* 736:255–261.
- Salas SP, Marshall G, Gutiérrez BL, Rosso P. (2006). Time course of maternal plasma volume and hormonal changes in women with preeclampsia or fetal growth restriction. *Hypertension* 47:203–208.
- Schönfelder G, Flick B, Mayr E, Talsness C, Paul M, Chahoud I. (2002a). In utero exposure to low doses of bisphenol A lead to long-term deleterious effects in the vagina. *Neoplasia* 2002;4:98–102.
- Schönfelder G, Wittfoht W, Hopp H, Talsness CE, Paul M, Chahoud I. (2002b). Parent bisphenol A accumulation in the human maternal-fetal-placental unit. *Environ Health Perspect* 110:A703–A707
- Sharpe RM. (2010). Is it time to end concerns over the estrogenic effects of bisphenol A? *Toxicol Sci* 114:1–4.
- Shin BS, Kim CH, Jun SY, Kim DW, Lee BM, Yoon CH, Park EH, Lee KC, Han S-Y, Park KL, Kim HS, Yoo SD. (2004). Physiologically based pharmacokinetics of bisphenol A. *J Toxicol Environ Health A* 67:1971–1985.
- Snyder RW, Maness SC, Gaido KW, Welsch F, Sumner SCJ, Fennell TR. (2000). Metabolism and disposition of bisphenol A in female rats. *Toxicol Appl Pharmacol* 168:225–234.
- Somm E, Schwitzgebel VM, Toulotte A, Cederroth CR, Combescure C, Nef S, Aubert ML, Hüppi PS. (2009). Perinatal exposure to bisphenol A alters early adipogenesis in the rat. *Environ Health Perspect* 117:1549–1555.
- Spivey A. (2009). Research issues and initiatives: NIEHS funds human BPA research. *Environ Health Perspect* 117:A541.
- Stahlhut RW, Welshons WV, Swan SH. Bisphenol A data in NHANES suggest longer than expected half-life, substantial nonfood exposure, or both. *Environ Health Perspect*. 2009;117:784–789.
- Stanley EL, Hume R, Coughtrie MW. (2005). Expression profiling of human fetal cytosolic sulfotransferases involved in steroid and thyroid hormone metabolism and in detoxification. *Mol Cell Endocrinol* 240:32–42.
- Stowell CL, Barvian KK, Young PC, Bigsby RM, Verdugo DE, Bertozzi CR, Widlanski TS. (2006). A role for sulfation-desulfation in the uptake of bisphenol a into breast tumor cells. *Chem Biol* 13:891–897.
- Stump DG, Beck MJ, Radovsky A, Garman RH, Freshwater LL, Sheets LP, Marty MS, Waechter JM Jr, Dimond SS, Van Miller JP, Shiotsuka RN, Beyer D, Chappelle AH, Hentges SG. (2010). Developmental neurotoxicity study of dietary bisphenol A in Sprague-Dawley rats. *Toxicol Sci* 115:167–182.
- Suiko M, Sakakibara Y, Liu MC. (2000). Sulfation of environmental estrogen-like chemicals by human cytosolic sulfotransferases. *Biochem Biophys Res Commun* 267:80–84.
- Takeuchi T, Tsutsumi O. (2002). Serum bisphenola concentrations showed gender differens, possibly linked to androgen levels. *Biochem Biophys Res Commun* 291:76–78.
- Tan E, Pang KS. (2001). Sulfation is rate limiting in the futile cycling between estrone and estrone sulphate in enriched periportal and perivenous rat hepatocytes. *Drug Metab Dispos* 29:335–346.
- Tan BL, Mustafa AM. (2003). Leaching of bisphenol A from new and old babies' bottles, and new babies' feeding teats. *Asia Pac J Public Health* 15:118–123.
- Taylor JA, vom Saal FS, Welshons WV, Drury B, Rottinghaus G, Hunt PA, VandeVoort CA. (2010). Similarity of bisphenol A pharmacokinetics in rhesus monkeys and mice: Relevance for human exposure. *Environ Health Perspect* [Online]. Accessed 20 September 2010. [Epub ahead of print].
- Teeguarden JG, Waechter JM Jr, Clewell HJ 3rd, Covington TR, Barton HA. (2005). Evaluation of oral and intravenous route pharmacokinetics, plasma protein binding, and uterine tissue dose metrics of bisphenol A: A physiologically based pharmacokinetic approach. *Toxicol Sci* 85:823–838.
- Thomson BM, Grounds PR. (2005). Bisphenol A in canned foods in New Zealand: An exposure assessment. *Food Addit Contam* 22:65–72.
- Timms BG, Howdeshell KL, Barton L, Bradley S, Richter CA, Vom Saal FS. (2005). Estrogenic chemicals in plastic and oral contraceptives disrupt development of the fetal mouse prostate and urethra. *Proc Natl Acad Sci U S A* 102:7014–7019.
- Tinwell H, Haseman J, Lefevre PA, Wallis N, Ashby J. (2002). Normal sexual development of two strains of rat exposed in utero to low doses of bisphenol A. *Toxicol Sci* 68:339–348.
- Tominaga T, Negishi T, Hirooka H, Miyachi A, Inoue A, Hayasaka I, Yoshikawa Y. (2006). Toxicokinetics of bisphenol A in rats, monkeys and chimpanzees by the LC-MS/MS method. *Toxicology* 226:208–217.
- Tsukioka T, Brock J, Graiser S, Nguyen J, Nakazawa H, Makino T. (2003). Determination of trace amounts of bisphenol A in urine by negative-ion chemical-ionization-gas chromatography/mass spectrometry. *Anal Sci* 19:151–153.
- Tsukioka T, Terasawa J, Sato S, Hatayama Y, Makino T, Nakazawa H. (2004). Development of analytical method for determining trace amounts of BPA in urine samples and estimation of exposure to BPA. *J Environ Chem* 14:57–63.
- Twaddle NC, Churchwell MI, Vanlandingham M, Doerge DR. (2010). Quantification of deuterated bisphenol A in serum, tissues, and excreta from adult Sprague-Dawley rats using liquid chromatography with tandem mass spectrometry. *Rapid Commun Mass Spectrom* 24:3011–3020.
- Tyl RW. (2003). Bisphenol A: Findings of a multigenerational rat study. *Environ Health Perspect* 111:A632.
- Tyl RW. (2009a). Basic exploratory research versus guideline-compliant studies used for hazard evaluation and risk assessment: Bisphenol A as a case study. *Environ Health Perspect* 117:1644–1651.
- Tyl RW (2009b). The presence (or not) of effects from low oral doses of BPA. *J Toxicol Sci* 34:587–588.
- Tyl RW, Myers CB, Marr MC, Thomas BE, Keimowitz AR, Brine DR, Veselica MM, Fail PA, Chang TY, Seely JC, Joiner RL, Butala JH, Dimond SS, Cagen SZ, Shiotsuka RN, Stropp GD, Waechter JM. (2002). Three-generation reproductive toxicity study of dietary bisphenol A in CD Sprague-Dawley rats. *Toxicol Sci* 68:121–146.
- Tyl RW, Myers CB, Marr MC, Fail PA, Seely JC, Elswick B, James A, Welsch F. (2003). Two-generation reproductive toxicity study of inhaled tertiary amyl methyl ether (TAME) vapor in CD rats. *J Appl Toxicol* 23:397–410.



- Tyl RW, Myers CB, Marr MC, Castillo NP, Veselica MM, Joiner RL, Dimond SS, Van Miller JP, Stropp GD, Waechter JM Jr, Hentges SG. (2008a). One-generation reproductive toxicity study of dietary 17beta-estradiol (E2; CAS No. 50-28-2) in CD-1 (Swiss) mice. *Reprod Toxicol* 25:144-160.
- Tyl RW, Myers CB, Marr MC, Sloan CS, Castillo NP, Veselica MM, Seely JC, Dimond SS, Van Miller JP, Shiotsuka RN, Beyer D, Hentges SG, Waechter JM Jr. (2008b). Two-generation reproductive toxicity study of dietary bisphenol A in CD-1 (Swiss) mice. *Toxicol Sci* 104:362-384.
- Tyl RW, Myers CB, Marr MC, Sloan CS, Castillo NP, Veselica MM, Seely JC, Dimond SS, Van Miller JP, Shiotsuka RS, Stropp GD, Waechter JM Jr, Hentges SG. (2008c). Two-generation reproductive toxicity evaluation of dietary 17beta-estradiol (E2; CAS No. 50-28-2) in CD-1 (Swiss) mice. *Toxicol Sci* 102:392-412.
- Uchida K, Suzuki A, Kobayashi K, Buchanan DL, Sato T, Watanabe H, Katsu Y, Suzuki J, Asaoka K, Mori C, Arizono K, Iguchi T. (2002). Bisphenol-A administration during pregnancy results in fetal exposure in mice and monkey. *J Health Sci* 48:579-582.
- Upmeier A, Degen GH, Diel P, Michna H, Bolt HM. (2000). Toxicokinetics of bisphenol A in female DA/Han rats after single i.v. and oral administration. *Arch Toxicol* 74:431-436.
- US EPA. (1984a). Ninety-day oral toxicity study in dogs. Office of Pesticides and Toxic Substances. Fiche No. OTS0509954.
- US EPA. (1984b). Reproduction and ninety-day oral toxicity study in rats. Office of Pesticides and Toxic Substances. Fiche No. OTS0509954.
- US EPA. (1984c). Fourteen-day range finding study in rats. Office of Pesticides and Toxic Substances. Fiche No. OTS0509954.
- USFDA. (2010). Update on bisphenol A for use in food contact applications. Available at: <http://www.fda.gov/NewsEvents/PublicHealthFocus/ucm197739.htm>, <http://www.fda.gov/downloads/NewsEvents/PublicHealthFocus/UCM197778.pdf>. (January 2010).
- US FDA Memorandum. (2009a). Summary of Bisphenol A Biomonitoring Studies. Komolprasert/Twaroski, 16/11/2009.
- US FDA Memorandum. (2009b). Bisphenol A (CAS RN. 80-05-7): Review of low dose studies. Aungst/Taroski, 08/31/2009.
- Valle LD, Toffolo V, Nardi A, Fiore C, Bernante P, Di Liddo R, Parnigotto PP, Colombo L. (2006). Tissue-specific transcriptional initiation and activity of steroid sulfatase complementing dehydroepiandrosterone sulfate uptake and intracrine steroid activations in human adipose tissue. *J Endocrinol* 190:129-139.
- Vandenberg LN, Hauser R, Marcus M, Olea N, Welshons WV. (2007). Human exposure to bisphenol A (BPA). *Reprod Toxicol* 24:139-177.
- Vandenberg LN, Maffini MV, Schaeberle CM, Ucci AA, Sonnenschein C, Rubin BS, Soto AM. (2008). Perinatal exposure to the xenoestrogen bisphenol-A induces mammary intraductal hyperplasias in adult CD-1 mice. *Reprod Toxicol* 26:210-219.
- Vandenberg LN, Chauhoud I, Heindel JJ, Padmanabhan V, Paumgartten FJ, Schoenfelder G. (2010a). Urinary, circulating and tissue biomonitoring studies indicate widespread exposure to bisphenol A. *Environ Health Perspect* 118:1055-1070.
- Vandenberg LN, Chahoud I, Padmanabhan V, Paumgartten FJ, Schoenfelder G. (2010b). Biomonitoring studies should be used by regulatory agencies to assess human exposure levels and safety of bisphenol A. *Environ Health Perspect* 118:1051-1054.
- Völkel W, Colnot T, Csanady G, Filser J, Dekant W. (2002). Metabolism and kinetics of bisphenol A in humans at low doses following oral administration. *Chem Res Toxicol* 15:1281-1287.
- Völkel W, Bittner N, Dekant W. (2005). Quantitation of bisphenol A and bisphenol A glucuronide in biological samples by high performance liquid chromatography-tandem mass spectrometry. *Drug Metabol Dispos* 33:1748-1757.
- Völkel W, Kiranoglu M, Fromme H. (2008). Determination of free and total bisphenol A in human urine to assess daily uptake as a basis for a valid risk assessment. *Toxicol Lett* 179:155-162.
- Vogel SA. (2009). The politics of plastics: The making and unmaking of bisphenol a "safety". *Am J Public Health* 99 (Suppl 3): S559-S566.
- Vom Saal FS, Myers JP. (2008). Bisphenol A and risk of metabolic disorders. *JAMA* 300:1353-1355.
- Vom Saal FS, Welshons WV. (2006). Large effects from small exposures. II. The importance of positive controls in low-dose research on bisphenol A. *Environ Res* 100:50-76.
- Waechter J Jr, Thornton C, Markham D, Domoradzki J. (2007). Factors affecting the accuracy of bisphenol A and bisphenol A-monoglucuronide estimates in mammalian tissues and urine samples. *Toxicol Mech Methods* 17:13-24.
- WHO. (2005). Chemical-specific adjustment factors for interspecies differences and human variability Guidance document for use of data in dose/concentration-response assessment. Geneva: World Health Organization.
- Willhite CC, Ball GL, McLellan CJ. (2008). Derivation of a bisphenol A oral reference dose (RfD) and drinking-water equivalent concentration. *J Toxicol Environ Health B Crit Rev* 11: 69-146.
- Wilson NK, Chuang JC, Morgan MK, Lordo RA, Sheldon LS. (2007). An observational study of the potential exposures of preschool children to pentachlorophenol, bisphenol-A, and nonylphenol at home and daycare. *Environ Res* 103:9-20.
- Yang M, Ryu JH, Jeon R, Kang D, Yoo KY. (2009). Effects of bisphenol A on breast cancer and its risk factors. *Arch Toxicol* 83:281-285.
- Ye X, Kuklenyik Z, Needham LL, Calafat AM. (2005). Quantification of urinary conjugates of bisphenol A, 2,5-dichlorophenol, and 2-hydroxy-4-methoxybenzophenone in humans by online solid phase extraction-high performance liquid chromatography-tandem mass spectrometry. *Anal Bioanal Chem* 383:638-644.
- Ye X, Bishop AM, Reidy JA, Needham LL, Calafat AM. (2007). Temporal stability of the conjugated species of bisphenol A, parabens, and other environmental phenols in human urine. *J Expo Sci Environ Epidemiol* 17:567-572.
- Ye X, Wong LY, Jia LT, Needham LL, Calafat AM. (2009). Stability of the conjugated species of environmental phenols and parabens in human serum. *Environ Int* 35:1160-1163.
- Zaya MJ, Hines RN, Stevens JC. (2006). Epirubicin glucuronidation and UGT2B7 developmental expression. *Drug Metab Dispos* 34:2097-2101.
- Zenser TV, Lakshmi VM, Davis BB. (1999). Human and *Escherichia coli* beta-glucuronidase hydrolysis of glucuronide conjugates of benzidine and 4-aminobiphenyl, and their hydroxy metabolites. *Drug Metab Dispos* 27:1064-1067.